


Original Russian text <https://vavilovj-icg.ru/>

# Candidate genes for domestication and resistance to cold climate according to whole genome sequencing data of Russian cattle and sheep breeds

N.S. Yudin<sup>1</sup>, D.M. Larkin<sup>2</sup> 

<sup>1</sup> Institute of Cytology and Genetics of the Siberian Branch of the Russian Academy of Sciences, Novosibirsk, Russia

<sup>2</sup> Royal Veterinary College, University of London, London, United Kingdom

 [dmlarkin@gmail.com](mailto:dmlarkin@gmail.com)

**Abstract.** It is known that different species of animals, when living in the same environmental conditions, can form similar phenotypes. The study of the convergent evolution of several species under the influence of the same environmental factor makes it possible to identify common mechanisms of genetic adaptation. Local cattle and sheep breeds have been formed over thousands of years under the influence of domestication, as well as selection aimed at adaptation to the local environment and meeting human needs. Previously, we identified a number of candidate genes in genome regions potentially selected during domestication and adaptation to the climatic conditions of Russia, in local breeds of cattle and sheep using whole genome genotyping data. However, these data are of low resolution and do not reveal most nucleotide substitutions. The aim of the work was to create, using the whole genome sequencing data, a list of genes associated with domestication, selection and adaptation in Russian cattle and sheep breeds, as well as to identify candidate genes and metabolic pathways for selection for cold adaptation. We used our original data on the search for signatures of selection in the genomes of Russian cattle (Yakut, Kholmogory, Buryat, Wagyu) and sheep (Baikal, Tuva) breeds. We used the HapFLK, DCMS, FST and PBS methods to identify DNA regions with signatures of selection. The number of candidate genes in potentially selective regions was 946 in cattle and 151 in sheep. We showed that the studied Russian cattle and sheep breeds have at least 10 genes in common, apparently involved in the processes of adaptation/selection, including adaptation to a cold climate, including the *ASTN2*, *PM20D1*, *TMEM176A*, and *GLIS1* genes. Based on the intersection with the list of selected genes in at least two Arctic/Antarctic mammal species, 20 and 8 genes, have been identified in cattle and sheep, respectively, that are potentially involved in cold adaptation. Among them, the most promising for further research are the *ASPH*, *NCKAP5L*, *SERPINF1*, and *SND1* genes. Gene ontology analysis indicated the existence of possible common biochemical pathways for adaptation to cold in domestic and wild mammals associated with cytoskeleton disassembly and apoptosis. Key words: signatures of selection; adaptation; cold; cattle; sheep; local breed; Russia; whole genome sequencing.


**For citation:** Yudin N.S., Larkin D.M. Candidate genes for domestication and resistance to cold climate according to whole genome sequencing data of Russian cattle and sheep breeds. *Vavilovskii Zhurnal Genetiki i Seleksii* = *Vavilov Journal of Genetics and Breeding*. 2023;27(5):463-470. DOI 10.18699/VJGB-23-56

## Гены-кандидаты доместикации и устойчивости к холоду по данным полногеномного секвенирования российских пород крупного рогатого скота и овец

Н.С. Юдин<sup>1</sup>, Д.М. Ларкин<sup>2</sup> 

<sup>1</sup> Федеральный исследовательский центр Институт цитологии и генетики Сибирского отделения Российской академии наук, Новосибирск, Россия

<sup>2</sup> Королевский Ветеринарный Колледж, Университет Лондона, Лондон, Великобритания

 [dmlarkin@gmail.com](mailto:dmlarkin@gmail.com)

**Аннотация.** Известно, что различные виды животных при обитании в одинаковых условиях среды могут сформировать сходные фенотипы. Изучение конвергентной эволюции нескольких видов под действием одного и того же средового фактора позволяет выявить у них общие механизмы генетической адаптации. Местные породы крупного и мелкого рогатого скота формировались на протяжении тысяч лет под воздействием доместикации, а также отбора, направленного на адаптацию к факторам местной среды обитания и удовлетворение потребностей человека. Ранее нами был выявлен ряд генов-кандидатов в участках генома, подвергшихся отбору в ходе доместикации и адаптации к климатическим условиям России, включая низкие зимние температуры, у местных пород крупного рогатого скота (КРС) и овец с использованием данных полногеномного секвенирования. Однако эти данные обладают низким разрешением и не позволяют выявить большинство нуклеотидных замен. Целью работы было создание по данным полногеномного секвенирования списка

генов, связанных с адаптацией российских пород КРС и овец, а также идентификация генов-кандидатов и метаболических путей для проведения селекции на адаптацию к холоду. Использованы опубликованные нами данные по поиску следов отбора в геномах российских или разводимых в России пород КРС (якутская, холмогорская, бурятская, вагю) и овец (забайкальская, тувинская). Количество генов-кандидатов в районах, потенциально подвергавшихся селекции, составило 946 у КРС и 151 у овец. Нами показано, что изученные российские породы КРС и овец имеют не менее 10 общих генов под отбором, по-видимому, участвующих в процессах адаптации/селекции, в том числе адаптации к холодному климату, включая гены *ASTN2*, *PM20D1*, *TMEM176A*, *GLIS1*. На основании пересечения со списком генов, подвергавшихся отбору по крайней мере у двух видов арктических/антарктических млекопитающих, у КРС и овец выявлено 20 и 8 генов соответственно, которые потенциально вовлечены в адаптацию к холоду. Среди них наиболее перспективными для дальнейших исследований являются *ASPH*, *NCKAP5L*, *SERPINF1* и *SND1*. Анализ геновых онтологий указывает на существование возможных общих биохимических путей адаптации к холоду у домашних и диких млекопитающих, связанных с разборкой цитоскелета и апоптозом.

Ключевые слова: признаки селекции; адаптация; холод; крупный рогатый скот; овца; местная порода; Россия; полногеномное секвенирование

## Introduction

The impact of extreme environmental factors can lead to either the extinction of a species or its adaptation to new environmental conditions (Nevo, 2011). It is known that different animal species, inhabiting similar conditions, can develop similar phenotypes using similar biochemical pathways (Storz, 2016). Studying convergent evolution of several species under the influence of the same environmental factor allows for the identification of common genetic adaptation mechanisms (Romashov et al., 2022).

For example, the same non-synonymous mutation in the rhodopsin gene independently arose and was subjected to selection in at least 20 species of fish in response to changes in water light conditions (Hill et al., 2019). The convergent amino acid substitutions also occurred in the prestin gene of whales and bats during the evolution of echolocation (Liu Y. et al., 2010). The non-synonymous substitution His207Arg in the melanocortin 1 receptor gene is associated with light feather coloration in red-footed boobies and ruffs (Lamichhaney et al., 2016). Our recent work on the northernmost cattle, the Yakut cattle from Siberia, revealed the phenomenon of convergent nucleotide evolution among domestic breeds and wild species living in similar harsh conditions and/or exhibiting similar phenotypes. We found the same amino acid substitution in the NRAP protein in Yakut cattle and 16 species of cold-adapted, hibernating or deep-diving mammals, which was absent in all other breeds of cattle and other Bovinae species in the "1000 Bull Genomes" dataset (Buggiotti et al., 2021). According to our data, this amino acid substitution presumably arose 500–800 years ago and is almost fixed in the modern Yakut cattle population.

It is known that the domestication of animals of different species is accompanied by a number of similar morphophysiological and behavioral changes (Belyaev, 1979; Wilkins et al., 2014). For example, one of the typical morphological features of domestication is the disruption of melanin synthesis, as well as a slowing down of melanocyte development, leading to the appearance of white spots on the body, up to the emergence of a uniform white color (Prasolova, Trut, 1993). Such phenotypic parallelism is observed in cattle, horses, pigs, dogs, cats, minks, chickens, pigeons, etc. (Larkin, Yudin,

2016). Indeed, when studying the genomes of populations of domestic animals, strong selection signals have been found in melanin metabolic pathway genes (*KIT*, *KITLG*, *MITF*, *PAX3*) (Cieslak et al., 2011).

Local breeds of cattle and sheep have been formed over thousands of years under the influence of domestication, as well as natural and artificial selection directed towards adaptation to the factors of the local environment and meeting human needs (Moiseeva et al., 2006; Kantanen et al., 2015). Studying the genomes of local breeds of cattle allows for the identification of genetic mechanisms of adaptation, including to low temperatures of the surrounding environment (Yudin et al., 2021). Earlier, we identified a number of candidate genes in genome regions that were potentially subject to selection during domestication and adaptation to harsh climatic conditions in Russia, in local breeds of cattle (*Bos taurus*) and sheep (*Ovis aries*) using data from whole-genome genotyping on standard SNP arrays (Yurchenko et al., 2018, 2019). Based on these results, we also identified 31 common candidate genes related to adaptation to the environment, including cold climate, in animals of the studied breeds (Yudin, Larkin, 2019). For example, the *NEB* gene, probably associated with heat production through shivering thermogenesis, was identified by us in genome regions subject to positive selection both in native Russian breeds of cattle and sheep, as well as in the genomes of the mammoth, polar bear, and minke whale.

However, whole-genome genotyping data have low resolution and do not allow the detection of most nucleotide substitutions in the genomes of different agricultural animal species. The aim of this study was to create a list of common genes associated with environmental adaptation in Russian breeds of cattle and sheep, as well as to identify promising genetic variants/candidate genes/metabolic pathways for further experiments, marker-assisted and genomic selection aimed at cold adaptation in agricultural animals, using whole-genome sequencing data. Previously, we analyzed selection signatures in the DNA samples from Yakut, Kholmogory, and Buryat cattle using GeneSeek Genomic Profiler High-Density SNP array containing approximately 139,000 SNPs (Yurchenko et al., 2018), and from Baikal and Tuva sheep using Ovine Infinium HD SNP BeadChip (Yurchenko et al., 2019).

## Materials and methods

In the study, we used our own published data on the search for selection signatures using whole-genome sequencing in the genomes of Russian or bred in Russia cattle breeds (Yakut, Kholmogory, Buryat, Wagyu) (Buggiotti et al., 2021; Igoshin et al., 2023) and sheep (Baikal, Tuva) (Sweet-Jones et al., 2021). High-throughput sequencing was performed in paired-end mode (150 bp + 150 bp) on the Illumina platform at Novogene Co., Ltd. (Hong Kong, China) for 20 animals per breed. The average coverage depth was at least 11x for cattle and 15x for sheep.

To identify regions potentially under selection pressure in the genomes of Buryat and Wagyu cattle, we used four complementary methods (Igoshin et al., 2023). The hapFLK method is based on statistics that consider haplotype structure in populations (Fariello et al., 2013). The DCMS method combines five whole-genome statistics: Fisher's fixation index (FST), haplotype homozygosity (H1), modified haplotype homozygosity (H12), Tajima's D index (D), and nucleotide diversity index (Pi) (Ma et al., 2015). The FST method identifies genome regions subject to selection by identifying DNA segments with high allelic frequency variability between compared populations (Porto-Neto et al., 2013). The PBS statistic uses pairwise FST values between three populations to quantitatively assess sequence differentiation (Yi et al., 2010). It is considered that genes with high differentiation between sequences may potentially be under positive selection. Candidate gene lists for further analysis of Buryat and Wagyu cattle were compiled by combining lists obtained by different methods. Potential selection regions in the genomes of Yakut and Kholmogory cattle breeds were identified using hapFLK statistics (Buggiotti et al., 2021). For the search for selection signatures in the genomes of Baikal and Tuva sheep, a computational pipeline based on the DCMS method was used (Yurchenko et al., 2019).

Gene identifiers in the Ensembl database were converted into gene symbols using the db2db tool ([http://biobnet.abcc.ncifcrf.gov/db/db2dbRes.php?input=inputType&outputs\[\]=outputType&idList=value\(s\)](http://biobnet.abcc.ncifcrf.gov/db/db2dbRes.php?input=inputType&outputs[]=outputType&idList=value(s))). Intersections between gene lists were analyzed using the Venn program (<http://bioinformatics.psb.ugent.be/webtools/Venn/>).

Biological functions of the shared genes that were under selection in Russian cattle and sheep breeds, and Arctic/Antarctic mammals were analyzed using the DAVID web tool (Huang et al., 2009). We identified enriched GO terms from the category of biological processes (GOTERM\_BP\_ALL) associated with four or more genes, compared to the control list of all human genes. We used a significance threshold criterion, characterized by the statistical significance of the observed number of genes with a specific GO term compared to the expected number of genes from the control list, and accepted  $p < 0.05$  as the threshold value.

## Results

The number of candidate genes in regions potentially subjected to selection was 946 for four Russian cattle breeds (*List\_Cattle*, Suppl. Material 1)<sup>1</sup> and 151 for two Russian sheep breeds (*List\_Sheep*, Suppl. Material 2) (see the Table). The

difference in the number of candidate genes between species is likely due to differences in the number of breeds included in the analysis, as well as the number of statistical methods used to detect signatures of selection (four statistics for Buryat and Wagyu cattle, one for the other breeds). Analysis of the intersection of the lists showed that 10 genes could potentially have been under selection in both species (see the Figure, *Cattle\_Sheep* list, Suppl. Material 3).

Previously, by intersecting the lists of genes potentially subjected to selection in six Arctic and Antarctic mammal species, we compiled a list of genes that may be involved in cold adaptation (Yudin et al., 2017). The list contained 416 genes that were likely under selection in at least two mammal species (*List\_Mammals*, Suppl. Material 4). To identify common genes that may be associated with adaptation to cold climate in Russian cattle and sheep breeds, we compared the lists of *List\_Mammals*, *List\_Cattle*, and *List\_Sheep*. As a result, we found 20 (*Cattle\_Mammals*, Suppl. Material 5) and 8 (*Sheep\_Mammals*, Suppl. Material 6) genes that were potentially under selection in at least two wild mammal species adapted to cold climate as well as in cattle and sheep, respectively (see the Figure).

To test the hypothesis that these lists were enriched in functional categories of genes related to cold adaptation, we performed gene ontology (GO) analysis on a list of 38 genes obtained by merging the *Cattle\_Sheep*, *Cattle\_Mammals*, and *Sheep\_Mammals* lists (*Cattle\_Sheep\_Mammals*, Suppl. Material 7). As a result, we found significant enrichment in eight GO terms that were associated with 4 or more genes (Suppl. Material 8).

## Discussion

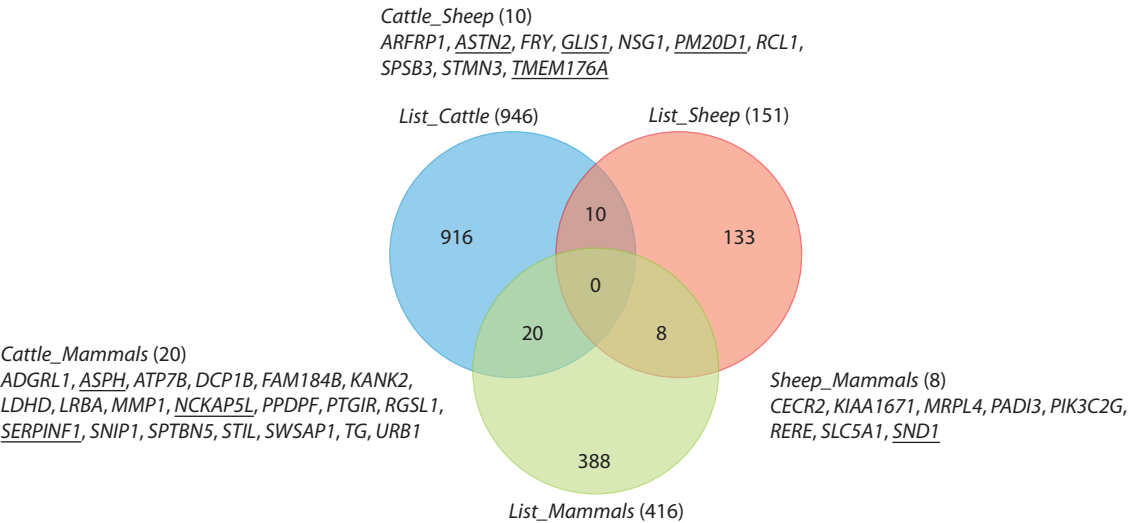
Our study aimed to identify common candidate genes in the genomes of domestic cattle and sheep breeds in Russia that may have undergone selection and played a role in adaptation to extreme climates, as well as to identify promising genetic variants/candidate genes/metabolic pathways for further cold adaptation research. We identified a total of 10 genes that potentially could have been under selection simultaneously in both Russian cattle and sheep breeds (*Cattle\_Sheep* list, see Suppl. Material 3). These genes were likely subjected to selection during domestication and/or subsequent selection for economically important traits, as well as during adaptation to cold climates.

According to the theory of D.K. Belyaev, numerous morphophysiological transformations in domestic animals are caused by destabilizing selection for the absence of aggressive behavior towards humans (Belyaev, 1979). Indeed, we have previously shown that a list of 1262 common genes that underwent selection in Russian cattle and sheep breeds through whole-genome genotyping was enriched in genes predominantly expressed in the brain (Yudin, Larkin, 2019). Several common genes identified in our study (see Suppl. Material 3) are expressed in nervous tissue and are involved in normal neuron function. For example, the protein astrotactin 2 (ASTN2) modulates synaptic activity in neurons by regulating the expression of synaptic proteins in post-migratory neurons via endocytosis (Behesti et al., 2018). Genetic variants in the *ASTN2* gene are associated with Alzheimer's disease (Wang et al., 2015), schizophrenia (Autism Spectrum Disorders Working

<sup>1</sup> Supplementary Materials 1–8 are available in the online version of the paper: [http://vavilov.elpub.ru/jour/manager/files/Suppl\\_Yudin\\_Engl\\_27\\_5.pdf](http://vavilov.elpub.ru/jour/manager/files/Suppl_Yudin_Engl_27_5.pdf)

Lists of potentially selected candidate genes

Name of the list in the Supplement	Description	Number of genes	Reference
List_Cattle (Suppl. Material 1)	List of genes potentially subjected to selection in the genomes of four Russian cattle breeds	946	Buggiotti et al., 2021; Igoshin et al., 2023
List_Sheep (Suppl. Material 2)	List of genes potentially subjected to selection in the genomes of two Russian sheep breeds	151	Sweet-Jones et al., 2021
Cattle_Sheep (Suppl. Material 3)	List of genes potentially subjected to selection in the genomes of Russian cattle and sheep breeds	10	–
List_Mammals (Suppl. Material 4)	List of genes potentially subjected to selection in the genomes of at least two species of Arctic/Antarctic mammals	416	Yudin et al., 2017
Cattle_Mammals (Suppl. Material 5)	List of genes potentially subjected to selection in the genomes of at least one Russian cattle breed, as well as at least two species of Arctic/Antarctic mammals	20	–
Sheep_Mammals (Suppl. Material 6)	List of genes potentially subjected to selection in the genomes of at least one Russian sheep breed, as well as at least two species of Arctic/Antarctic mammals	8	–
Cattle_Sheep_Mammals (Suppl. Material 7)	List of genes potentially subjected to selection in the genomes of Russian cattle, sheep, as well as Arctic/Antarctic mammals – combination of the Cattle_Sheep, Cattle_Mammals, and Sheep_Mammals lists	38	–



Venn diagram showing overlaps between lists of genes potentially subjected to selection in the genomes of Russian cattle (*List\_Cattle*) and sheep (*List\_Sheep*) breeds, and at least two species of Arctic/Antarctic mammals (*List\_Mammals*). The number of genes in each list is indicated in parentheses. The most promising cold adaptation candidate genes (based on their biological role) are underlined.

Group of The Psychiatric Genomics Consortium, 2017), autism (Lionel et al., 2014), and other psychiatric disorders. The gene encoding the protein containing a peptidase domain M20 1 (*PM20D1*) is associated with Alzheimer’s disease (Sanchez-Mut et al., 2018) and Parkinson’s disease (Rudakou et al., 2021). The transmembrane protein *TMEM176A* gene is associated with schizophrenia (Kos et al., 2017). At the same time, deletion in the *ASTN2* gene results in a reversal of normal orientation of hair follicles in adult mice (parallel to “from head to tail”) to the opposite direction

(parallel to “from tail to head”) (Chang et al., 2015). In humans, the *ASTN2* gene is associated with the level of triglycerides in the blood (Jiao et al., 2015) and the development of obesity (Burt et al., 2021). Signatures of selection in this gene have been found in ethnic groups of southern Ethiopia, who have lived in high-altitude conditions for over a thousand years (Scheinfeldt et al., 2012). Interestingly, adaptive introgression of a large number of ancient Neanderthal alleles has been identified in the *ASTN2* gene in the population of South Asia (Racimo et al., 2017). The biochemical pathway of *PM20D1*



modulates the accumulation of brown fat and thus participates in the process of heat production through non-shivering thermogenesis (Gao et al., 2018). The pro-adipogenic factor GLIS1 may play a critical role in the differentiation of mesodermal cells during fetal development and affect fat distribution in the tail of sheep (Luo et al., 2021). SNP polymorphism in the *NSG1* gene is associated with the fat content in milk of Holstein cows (Lee et al., 2016).

We investigated promising genetic variants and candidate genes for cold adaptation by intersecting the lists *List\_Cattle* and *List\_Sheep* with the *List\_Mammals* of 416 genes that were positively selected in at least two species of Arctic/Antarctic mammals (Yudin et al., 2017). When all three lists of common genes and genetic variants were intersected, none were found (see the Figure), but we identified 20 and 8 genes (see Suppl. Materials 5 and 6), respectively, that were potentially subjected to selection during adaptation to the climate of both Arctic mammals and cattle or sheep, respectively.

Thus, genetic variants in the *ASPH* gene, which encodes a protein that regulates the process of excitation–contraction in muscles, are associated with heat stroke and malignant hyperthermia in humans (Endo et al., 2022). According to whole-genome association analysis, single nucleotide polymorphisms in this gene are associated with intramuscular fat distribution in beef cattle (Ramayo-Caldas et al., 2014). Genetic variants in the *NCKAP5L* (Chen et al., 2013) and *SERPINF1* (Böhm et al., 2012) genes are associated with the development of obesity in humans. The human gene *FAM184B* is associated with body composition and fatty acid profile (Yuan et al., 2021). The protein product of the *PADI3* gene controls hair shape on the human scalp (Liu F. et al., 2018). The mRNA expression of the gene of the protein containing the staphylococcal nuclease domain 1 (*SND1*) in the New Zealand alpine stick insect significantly increases in response to cold exposure (Dunning et al., 2013). In mammals, *SND1* plays an important role in regulating lipid metabolism through the activation of the *SREBP2* protein (Navarro-Imaz et al., 2020).

The gene ontology terms identified by the DAVID program when analyzing the list of potentially selected genes in Russian cattle, sheep, as well as Arctic/Antarctic mammals (*Cattle\_Sheep\_Mammals* list) can be divided into three groups: (1) terms related to the disassembly of cell parts and protein complexes (“disassembly of cell components”, “disassembly of protein complex”, “disassembly of macromolecular complex”, etc.); (2) terms related to DNA disintegration (“hydrolysis of phosphodiester bonds in nucleic acids”); (3) uninformative terms of the top hierarchy describing general biological processes (“biological process occurring at the level of a multicellular organism”) (see Suppl. Material 8).

Enrichment of gene ontology terms related to the disassembly of cell parts, proteins, and DNA may be the result of natural selection for genes encoding cytoskeletal proteins and/or participating in programmed cell death (apoptosis). Studies on hibernating mammals have shown that their cells respond to low temperatures by disassembling the cytoskeleton and delaying apoptosis (Van Breukelen, Martin, 2002). It is believed that cytoskeletal disassembly may be the cause of protein synthesis suppression in mammalian cells during cold stress (Al-Fageeh, Smales, 2006). Hypothermia

causes disassembly of microtubules by activating p38 MAP kinase in human retinal cells (Thanuja et al., 2021). In *in vivo* and *in vitro* experiments, it has been shown that microtubules in peripheral axons of *Xenopus* are sensitive to cold, and their density varies depending on the season (Alvarez, Fadić, 1992). It has been shown that cold stress induces apoptosis of neurons in the hippocampus of mice (Xu et al., 2019).

In our study, the term “disassembly of cellular components” was associated with seven genes (see Suppl. Material 8). Among them, the gene *SPTBN5* encodes one of the spectrin family proteins, which are common components of the cytoskeleton, interacting with elements of the cell scaffold and plasma membrane, providing proper localization of major membrane proteins, signal transmission into the cell, and other processes (Beijer, Züchner, 2022). The protein *NCKAP5L*, interacting with the protein *CDK5RAP2*, regulates microtubule stability in HeLa cells (Mori et al., 2015). The protein stathmin-3, encoded by the *STMN3* gene, regulates the rapid reorganization of the cytoskeleton in response to environmental factors by affecting the balance of microtubule assembly and disassembly (Nair et al., 2014). With the term “hydrolysis of phosphodiester bonds of nucleic acids”, genes *RCL1*, *CECR2*, *SND1*, and *DCPIB* were associated (see Suppl. Material 8). It has been shown that the *SND1* protein suppresses apoptosis in hepatocellular carcinoma cells by interacting with the long non-coding RNA *UCA1* (Cui et al., 2018). The *CECR2* protein is localized in the DNA condensation regions of apoptotic human liver cells and interacts with the chromatin-associated protein *TAFII30* (Liu L. et al., 2002).

## Conclusion

Thus, using whole-genome sequencing data, we have shown that the studied Russian cattle and sheep breeds have at least 10 common genes, presumably involved in adaptation/selection processes, including adaptation to cold climate, such as *ASTN2*, *PM20D1*, *TMEM176A*, *GLIS1*. Based on the overlap with the list of genes subjected to selection in at least two species of Arctic/Antarctic mammals, 20 and 8 genes potentially involved in adaptation to cold were identified in cattle and sheep, respectively. Among them, the most promising for further research are the genes *ASPH*, *NCKAP5L*, *SERPINF1*, and *SND1*. Gene ontology analysis indicates the existence of possible common biochemical pathways for adaptation to cold in domestic and wild mammals, related to cytoskeleton disassembly and apoptosis.

## References

- Al-Fageeh M.B., Smales C.M. Control and regulation of the cellular responses to cold shock: the responses in yeast and mammalian systems. *Biochem. J.* 2006;397(2):247-259. DOI 10.1042/BJ20060166.
- Alvarez J., Fadić R. Assembly and disassembly of axonal microtubules of the toad *Xenopus laevis* under the effect of temperature. *J. Exp. Zool.* 1992;264(3):261-266. DOI 10.1002/jez.1402640305.
- Autism Spectrum Disorders Working Group of The Psychiatric Genomics Consortium. Meta-analysis of GWAS of over 16,000 individuals with autism spectrum disorder highlights a novel locus at 10q24.32 and a significant overlap with schizophrenia. *Mol. Autism.* 2017;8:21. DOI 10.1186/s13229-017-0137-9.

- Behesti H., Fore T.R., Wu P., Horn Z., Leppert M., Hull C., Hat-ten M.E. ASTN2 modulates synaptic strength by trafficking and degradation of surface proteins. *Proc. Natl. Acad. Sci. USA*. 2018;115(41):E9717-E9726. DOI 10.1073/pnas.1809382115.
- Beijer D., Züchner S.L. Commentary: *SPTBN5*, encoding the  $\beta$ V-spectrin protein, leads to a syndrome of intellectual disability, developmental delay, and seizures. *Front. Mol. Neurosci*. 2022;15:1011856. DOI 10.3389/fnmol.2022.1011856.
- Belyaev D.K. The Wilhelmine E. Key 1978 invitational lecture. Destabilizing selection as a factor in domestication. *J. Hered*. 1979;70(5):301-308. DOI 10.1093/oxfordjournals.jhered.a109263.
- Böhm A., Ordelheide A.M., Machann J., Heni M., Ketterer C., Machicao F., Schick F., Stefan N., Fritsche A., Häring H.U., Staiger H. Common genetic variation in the *SERPINF1* locus determines overall adiposity, obesity-related insulin resistance, and circulating leptin levels. *PLoS One*. 2012;7(3):e34035. DOI 10.1371/journal.pone.0034035.
- Buggiotti L., Yurchenko A.A., Yudin N.S., Vander Jagt C.J., Vorobieva N.V., Kusliy M.A., Vasiliev S.K., Rodionov A.N., Boronetskaya O.I., Zinovieva N.A., Graphodatsky A.S., Daetwyler H.D., Larkin D.M. Demographic history, adaptation, and NRAP convergent evolution at amino acid residue 100 in the world northernmost cattle from Siberia. *Mol. Biol. Evol*. 2021;38(8):3093-3110. DOI 10.1093/molbev/msab078.
- Burt O., Johnston K.J.A., Graham N., Cullen B., Lyall D.M., Lyall L.M., Pell J.P., Ward J., Smith D.J., Strawbridge R.J. Genetic variation in the ASTN2 locus in cardiovascular, metabolic and psychiatric traits: evidence for pleiotropy rather than shared biology. *Genes (Basel)*. 2021;12(8):1194. DOI 10.3390/genes12081194.
- Chang H., Cahill H., Smallwood P.M., Wang Y., Nathans J. Identification of Astrotactin2 as a genetic modifier that regulates the global orientation of mammalian hair follicles. *PLoS Genet*. 2015;11(9):e1005532. DOI 10.1371/journal.pgen.1005532.
- Chen J., Meng Y., Zhou J., Zhuo M., Ling F., Zhang Y., Du H., Wang X. Identifying candidate genes for Type 2 Diabetes Mellitus and obesity through gene expression profiling in multiple tissues or cells. *J. Diabetes Res*. 2013;2013:970435. DOI 10.1155/2013/970435.
- Cieslak M., Reissmann M., Hofreiter M., Ludwig A. Colours of domestication. *Biol. Rev. Camb. Philos. Soc*. 2011;86(4):885-899. DOI 10.1111/j.1469-185X.2011.00177.x.
- Cui X., Zhao C., Yao X., Qian B., Su C., Ren Y., Yao Z., Gao X., Yang J. SND1 acts as an anti-apoptotic factor via regulating the expression of lncRNA *UCAL1* in hepatocellular carcinoma. *RNA Biol*. 2018;15(10):1364-1375. DOI 10.1080/15476286.2018.1534525.
- Dunning L.T., Dennis A.B., Park D., Sinclair B.J., Newcomb R.D., Buckley T.R. Identification of cold-responsive genes in a New Zealand alpine stick insect using RNA-Seq. *Comp. Biochem. Physiol. Part D Genomics Proteomics*. 2013;8(1):24-31. DOI 10.1016/j.cbd.2012.10.005.
- Endo Y., Groom L., Celik A., Kraeva N., Lee C.S., Jung S.Y., Gardner L., Shaw M.A., Hamilton S.L., Hopkins P.M., Dirksen R.T., Riaz S., Dowling J.J. Variants in *ASPH* cause exertional heat illness and are associated with malignant hyperthermia susceptibility. *Nat. Commun*. 2022;13(1):3403. DOI 10.1038/s41467-022-31088-8.
- Fariello M.I., Boitard S., Naya H., SanCristobal M., Servin B. Detecting signatures of selection through haplotype differentiation among hierarchically structured populations. *Genetics*. 2013;193(3):929-941. DOI 10.1534/genetics.112.147231.
- Gao Y., Qimuge N.R., Qin J., Cai R., Li X., Chu G.Y., Pang W.J., Yang G.S. Acute and chronic cold exposure differentially affects the browning of porcine white adipose tissue. *Animal*. 2018;12(7):1435-1441. DOI 10.1017/S1751731117002981.
- Hill J., Enbody E.D., Pettersson M.E., Sprehn C.G., Bekkevold D., Folkvord A., Laikre L., Kleinau G., Scheerer P., Andersson L. Recurrent convergent evolution at amino acid residue 261 in fish rhodopsin. *Proc. Natl. Acad. Sci. USA*. 2019;116(37):18473-18478. DOI 10.1073/pnas.1908332116.
- Huang D.W., Sherman B.T., Lempicki R.A. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat. Protoc*. 2009;4(1):44-57. DOI 10.1038/nprot.2008.211.
- Igoshin A.V., Romashov G.A., Yurchenko A.A., Yudin N.S., Larkin D.M. Resequencing commercial and locally adapted Turano-Mongolian cattle breeds reveals differences in selective pressures. *Animal Genetics*. 2023 (Submitted).
- Jiao H., Wang K., Yang F., Grant S.F., Hakonarson H., Price R.A., Li W.D. Pathway-based genome-wide association studies for plasma triglycerides in obese females and normal-weight controls. *PLoS One*. 2015;10(8):e0134923. DOI 10.1371/journal.pone.0134923.
- Kantanen J., Lovendahl P., Strandberg E., Eythorsdottir E., Li M.H., Kettunen-Praebel A., Berg P., Meuwissen T. Utilization of farm animal genetic resources in a changing agro-ecological environment in the Nordic countries. *Front. Genet*. 2015;6:52. DOI 10.3389/fgene.2015.00052.
- Kos M.Z., Carless M.A., Peralta J., Curran J.E., Quillen E.E., Almeida M., Blackburn A., Blondell L., Roalf D.R., Pogue-Geile M.F., Gur R.C., Göring H.H.H., Nimgaonkar V.L., Gur R.E., Almasy L. Exome sequences of multiplex, multigenerational families reveal schizophrenia risk loci with potential implications for neurocognitive performance. *Am. J. Med. Genet. B Neuropsychiatr. Genet*. 2017;174(8):817-827. DOI 10.1002/ajmg.b.32597.
- Lamichhane S., Fan G., Widemo F., Gunnarsson U., Thalmann D.S., Hoepfner M.P., Kerje S., Gustafson U., Shi C., Zhang H., Chen W., Liang X., Huang L., Wang J., Liang E., Wu Q., Lee S.M., Xu X., Höglund J., Liu X., Andersson L. Structural genomic changes underlie alternative reproductive strategies in the ruff (*Philomachus pugnax*). *Nat. Genet*. 2016;48(1):84-88. DOI 10.1038/ng.3430.
- Larkin D.M., Yudin N.S. The genomes and history of domestic animals. *Mol. Genet. Microbiol. Virol*. 2016;31(4):197-202. DOI 10.3103/S0891416816040054.
- Lee Y.S., Shin D., Lee W., Taye M., Cho K., Park K.D., Kim H. The prediction of the expected current selection coefficient of single nucleotide polymorphism associated with Holstein milk yield, fat and protein contents. *Asian-Australas. J. Anim. Sci*. 2016;29(1):36-42. DOI 10.5713/ajas.15.0476.
- Lionel A.C., Tammimies K., Vaags A.K., Rosenfeld J.A., Ahn J.W., Merico D., Noor A., Runke C.K., Pillalamarri V.K., Carter M.T., ... Ogilvie C.M., Shen Y., Hodge J.C., Talkowski M.E., Stavropoulos D.J., Marshall C.R., Scherer S.W. Disruption of the *ASTN2/TRIM32* locus at 9q33.1 is a risk factor in males for autism spectrum disorders, ADHD and other neurodevelopmental phenotypes. *Hum. Mol. Genet*. 2014;23(10):2752-2768. DOI 10.1093/hmg/ddt669.
- Liu F., Chen Y., Zhu G., Hysi P.G., Wu S., Adhikari K., Breslin K., Pospiech E., Hamer M.A., Peng F., Muralidharan C., Acuna-Alonzo V., Canizales-Quinteros S., Bedoya G., Gallo C., Poletti G., Rothhammer F., Bortolini M.C., Gonzalez-Jose R., Zeng C., Xu S., Jin L., Uitterlinden A.G., Ikram M.A., van Duijn C.M., Nijsten T., Walsh S., Branicki W., Wang S., Ruiz-Linares A., Spector T.D., Martin N.G., Medland S.E., Kayser M. Meta-analysis of genome-wide association studies identifies 8 novel loci involved in shape variation of human head hair. *Hum. Mol. Genet*. 2018;27(3):559-575. DOI 10.1093/hmg/ddx416.
- Liu L., Amy V., Liu G., McKeehan W.L. Novel complex integrating mitochondria and the microtubular cytoskeleton with chro-

- mosome remodeling and tumor suppressor RASSF1 deduced by in silico homology analysis, interaction cloning in yeast, and colocalization in cultured cells. *In Vitro Cell. Dev. Biol. Anim.* 2002;38(10):582-594. DOI 10.1290/1543-706x(2002)38<582:ncimat>2.0.co;2.
- Liu Y., Cotton J.A., Shen B., Han X., Rossiter S.J., Zhang S. Convergent sequence evolution between echolocating bats and dolphins. *Curr. Biol.* 2010;20(2):R53-R54. DOI 10.1016/j.cub.2009.11.058.
- Luo R., Zhang X., Wang L., Zhang L., Li G., Zheng Z. *GLIS1*, a potential candidate gene affect fat deposition in sheep tail. *Mol. Biol. Rep.* 2021;48(5):4925-4931. DOI 10.1007/s11033-021-06468-w.
- Ma Y., Ding X., Qanbari S., Weigend S., Zhang Q., Simianer H. Properties of different selection signature statistics and a new strategy for combining them. *Heredity (Edinb.)*. 2015;115(5):426-436. DOI 10.1038/hdy.2015.42.
- Moiseeva I.G., Ukhonov S.V., Stolpovsky Yu.A., Sulimova G.E., Kashtanov S.N. Gene Pools of Farm Animals. Livestock Genetic Resources in Russia. Moscow: Nauka Publ., 2006. (in Russian)
- Mori Y., Inoue Y., Tanaka S., Doda S., Yamanaka S., Fukuchi H., Terada Y. Cep169, a novel microtubule plus-end-tracking centrosomal protein, binds to CDK5RAP2 and regulates microtubule stability. *PLoS One*. 2015;10(10):e0140968. DOI 10.1371/journal.pone.0140968.
- Nair S., Bora-Singhal N., Perumal D., Chellappan S. Nicotine-mediated invasion and migration of non-small cell lung carcinoma cells by modulating *STMN3* and *GSPT1* genes in an ID1-dependent manner. *Mol. Cancer*. 2014;13:173. DOI 10.1186/1476-4598-13-173.
- Navarro-Imaz H., Ochoa B., García-Arcos I., Martínez M.J., Chico Y., Fresnedo O., Rueda Y. Molecular and cellular insights into the role of *SND1* in lipid metabolism. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids*. 2020;1865(5):158589. DOI 10.1016/j.bbalip.2019.158589.
- Nevo E. Evolution under environmental stress at macro- and micro-scales. *Genome Biol. Evol.* 2011;3:1039-1052. DOI 10.1093/gbe/evr052.
- Porto-Neto L.R., Lee S.H., Lee H.K., Gondro C. Detection of signatures of selection using  $F_{ST}$ . In: Gondro C., van der Werf J., Hayes B. (Eds.) *Genome-Wide Association Studies and Genomic Prediction. Methods in Molecular Biology*. Totowa, NJ: Humana Press, 2013;1019:423-436. DOI 10.1007/978-1-62703-447-0\_19.
- Prasolova L.A., Trut L.N. The effect of the *Star* gene on the rate of melanoblast migration in silver fox embryos. *Doklady RAN = Proceedings of the Russian Academy of Sciences*. 1993;329(6):787-789. (in Russian)
- Racimo F., Marnetto D., Huerta-Sánchez E. Signatures of archaic adaptive introgression in present-day human populations. *Mol. Biol. Evol.* 2017;34(2):296-317. DOI 10.1093/molbev/msw216.
- Ramayo-Caldas Y., Fortes M.R., Hudson N.J., Porto-Neto L.R., Bolormaa S., Barendse W., Kelly M., Moore S.S., Goddard M.E., Lehnert S.A., Reverter A. A marker-derived gene network reveals the regulatory role of *PPARGC1A*, *HNF4G*, and *FOXP3* in intramuscular fat deposition of beef cattle. *J. Anim. Sci.* 2014;92(7):2832-2845. DOI 10.2527/jas.2013-7484.
- Romashov G., Yurchenko A., Yudin N., Larkin D. How does a harsh environment make wild and domestic animals evolve similarly? In: Veerkamp R.F., de Haas Y. (Eds.) *Proceedings of 12th World Congress on Genetics Applied to Livestock Production (WCGALP)*. Rotterdam, The Netherlands, 3–8 July 2022. Wageningen: Wageningen Acad. Publishers, 2022;1078-1081. DOI 10.3920/978-90-8686-940-4\_255.
- Rudakou U., Yu E., Krohn L., Ruskey J.A., Asayesh F., Dauviliers Y., Spiegelman D., Greenbaum L., Fahn S., Waters C.H., Dupré N., Rouleau G.A., Hassin-Baer S., Fon E.A., Alcalay R.N., Gan-Or Z. Targeted sequencing of Parkinson's disease loci genes highlights *SYT11*, *FGF20* and other associations. *Brain*. 2021;144(2):462-472. DOI 10.1093/brain/awaa401.
- Sanchez-Mut J.V., Heyn H., Silva B.A., Dixsaut L., Garcia-Esparcia P., Vidal E., Sayols S., Glauser L., Monteagudo-Sánchez A., Perez-Tur J., Ferrer I., Monk D., Schneider B., Esteller M., Gräff J. *PM20D1* is a quantitative trait locus associated with Alzheimer's disease. *Nat. Med.* 2018;24(5):598-603. DOI 10.1038/s41591-018-0013-y.
- Scheinfeldt L.B., Soi S., Thompson S., Ranciaro A., Woldemeskel D., Beggs W., Lambert C., Jarvis J.P., Abate D., Belay G., Tishkoff S.A. Genetic adaptation to high altitude in the Ethiopian highlands. *Genome Biol.* 2012;13(1):R1. DOI 10.1186/gb-2012-13-1-r1.
- Storz J.F. Causes of molecular convergence and parallelism in protein evolution. *Nat. Rev. Genet.* 2016;17(4):239-250. DOI 10.1038/nrg.2016.11.
- Sweet-Jones J., Yurchenko A.A., Igoshin A.V., Yudin N.S., Swain M.T., Larkin D.M. Resequencing and signatures of selection scan in two Siberian native sheep breeds point to candidate genetic variants for adaptation and economically important traits. *Anim. Genet.* 2021;52(1):126-131. DOI 10.1111/age.13015.
- Thanuja M.Y., Suma B.S., Dinesh D., Ranganath S.H., Srinivas S.P. Microtubule stabilization protects hypothermia-induced damage to the cytoskeleton and barrier integrity of the corneal endothelial cells. *J. Ocul. Pharmacol. Ther.* 2021;37(7):399-411. DOI 10.1089/jop.2021.0036.
- Van Breukelen F., Martin S.L. Invited review: molecular adaptations in mammalian hibernators: unique adaptations or generalized responses? *J. Appl. Physiol.* 2002;92(6):2640-2647. DOI 10.1152/japplphysiol.01007.2001.
- Wang K.S., Tonarelli S., Luo X., Wang L., Su B., Zuo L., Mao C., Rubin L., Briones D., Xu C. Polymorphisms within *ASTN2* gene are associated with age at onset of Alzheimer's disease. *J. Neural. Transm. (Vienna)*. 2015;122(5):701-708. DOI 10.1007/s00702-014-1306-z.
- Wilkins A.S., Wrangham R.W., Fitch W.T. The "domestication syndrome" in mammals: a unified explanation based on neural crest cell behavior and genetics. *Genetics*. 2014;197(3):795-808. DOI 10.1534/genetics.114.165423.
- Xu B., Lang L.M., Li S.Z., Guo J.R., Wang J.F., Wang D., Zhang L.P., Yang H.M., Lian S. Cortisol excess-mediated mitochondrial damage induced hippocampal neuronal apoptosis in mice following cold exposure. *Cells*. 2019;8(6):612. DOI 10.3390/cells8060612.
- Yi X., Liang Y., Huerta-Sánchez E., Jin X., Cuo Z.X., Pool J.E., Xu X., Jiang H., Vinckenbosch N., Korneliussen T.S., ... Zhang X., Li R., Li S., Yang H., Nielsen R., Wang Jun, Wang Jian. Sequencing of 50 human exomes reveals adaptation to high altitude. *Science*. 2010;329(5987):75-78. DOI 10.1126/science.1190371.
- Yuan Z., Sunduimijid B., Xiang R., Behrendt R., Knight M.I., Mason B.A., Reich C.M., Prowse-Wilkins C., Vander Jagt C.J., Chamberlain A.J., MacLeod I.M., Li F., Yue X., Daetwyler H.D. Expression quantitative trait loci in sheep liver and muscle contribute to variations in meat traits. *Genet. Sel. Evol.* 2021;53(1):8. DOI 10.1186/s12711-021-00602-9.
- Yudin N.S., Larkin D.M. Shared signatures of selection related to adaptation and acclimation in local cattle and sheep breeds from Russia. *Russ. J. Genet.* 2019;55(8):1008-1014. DOI 10.1134/S1022795419070159.
- Yudin N.S., Larkin D.M., Ignatieva E.V. A compendium and functional characterization of mammalian genes involved in adaptation to Arctic or Antarctic environments. *BMC Genet.* 2017;18(Suppl.1):111. DOI 10.1186/s12863-017-0580-9.

Yudin N.S., Yurchenko A.A., Larkin D.M. Signatures of selection and candidate genes for adaptation to extreme environmental factors in the genomes of Turano-Mongolian cattle breeds. *Vavilovskii Zhurnal Genetiki i Seleksii* = *Vavilov Journal of Genetics and Breeding*. 2021;25(2):190-201. DOI 10.18699/VJ21.023. (in Russian)

Yurchenko A.A., Daetwyler H.D., Yudin N., Schnabel R.D., Vander Jagt C.J., Soloshenko V., Lhasaranov B., Popov R., Taylor J.F., Larkin D.M. Scans for signatures of selection in Russian cattle breed genomes reveal new candidate genes for environmental

adaptation and acclimation. *Sci. Rep.* 2018;8(1):12984. DOI 10.1038/s41598-018-31304-w.

Yurchenko A.A., Deniskova T.E., Yudin N.S., Dotsev A.V., Khamiruev T.N., Selionova M.I., Egorov S.V., Reyer H., Wimmers K., Brem G., Zinovieva N.A., Larkin D.M. High-density genotyping reveals signatures of selection related to acclimation and economically important traits in 15 local sheep breeds from Russia. *BMC Genomics*. 2019;20(Suppl.3):294. DOI 10.1186/s12864-019-5537-0.

---

#### ORCID ID

Yudin N.S. [orcid.org/0000-0002-1947-5554](https://orcid.org/0000-0002-1947-5554)

Larkin D.M. [orcid.org/0000-0001-7859-6201](https://orcid.org/0000-0001-7859-6201)

**Acknowledgements.** This work was funded by the Russian Scientific Foundation (RSF) grant No. 19-76-20026.

**Conflict of interest.** The authors declare no conflict of interest.

Received December 2, 2022. Revised February 19, 2023. Accepted February 20, 2023.