

Prospects for marker-associated selection in tomato *Solanum lycopersicum* L.

A.B. Shcherban

Institute of Cytology and Genetics, SB RAS, Novosibirsk, Russia
✉ e-mail: atos@bionet.nsc.ru

The review gives a brief description of tomato, one of the main objects of olericulture for Siberia. The data on the main directions in the breeding of this culture, such as resistance to various pathogens, the nutritional properties of fruits, the timing of their maturation and storage are generalized. A separate chapter is devoted to the use of various types of DNA markers for constructing detailed genetic maps of the specified object, which, along with full-genome sequencing data, can be used to screen for genes responsible for breeding traits. Most of these traits, especially specific resistance to one or another pathogen, were transferred to the cultivated tomato by crossing with wild species, therefore, special attention was paid in the article to identifying and marking resistance genes to a variety of viral, fungal and bacterial pathogens occurring in Western Siberia and adjacent areas. Another important aspect for breeding is the nutrient content of tomato fruits, including carotenoids, vitamins, sugars, organic acids, etc. Recently, due to modern technologies of sequencing, SNP-genotyping, the development of new bioinformatic approaches, it has become possible to establish genetic cascades determining the biochemical composition of tomato fruits, to identify key genes that can be used in the future for marker-associated selection of nutritional value. And, finally, genetic works devoted to the problem of the optimal dates of fruit ripening in certain climatic conditions and their prolonged storage without loss of quality are discussed.

Key words: tomato; selection; DNA marker; pathogen; resistance; ripening time; shelf life.

For citation: Shcherban A.B. Prospects for marker-associated selection in tomato *Solanum lycopersicum* L. Vavilovskii Zhurnal Genetiki i Selektzii = Vavilov Journal of Genetics and Breeding. 2019;23(5):534-541. DOI 10.18699/VJ19.522

Перспективы маркер-ориентированной селекции томата *Solanum lycopersicum* L.

А.Б. Щербань

Федеральный исследовательский центр Институт цитологии и генетики Сибирского отделения Российской академии наук, Новосибирск, Россия
✉ e-mail: atos@bionet.nsc.ru

В обзоре представлена краткая характеристика одного из основных для Сибири объектов овощеводства – томата. Обобщены данные об основных направлениях селекции этой культуры, таких как устойчивость к различным патогенам, сроки созревания и хранения плодов, а также содержание в них биологически активных веществ (БАВ). Отдельная глава обзора посвящена использованию различных типов маркеров ДНК для построения детальных генетических карт указанного объекта, которые наряду с данными полногеномного секвенирования могут быть использованы для скрининга различных генов, отвечающих за селективируемые признаки. Большинство таких признаков, особенно специфическая устойчивость к тем или иным патогенам, перенесено в культурный томат путем скрещивания его с дикорастущими видами, поэтому особое внимание в статье уделено выявлению и маркированию генов устойчивости к целому ряду вирусных, грибных и бактериальных патогенов, распространенных в Западной Сибири и на прилегающих территориях. Другой важный аспект для селекции – содержание БАВ в плодах томата, включая каротиноиды, витамины, сахара, органические кислоты и др. За последнее время благодаря современным технологиям секвенирования, SNP-генотипирования, разработке новых биоинформатических подходов удалось установить генетические каскады, определяющие биохимический состав плодов томата; выделить ключевые гены, которые в перспективе могут быть использованы в маркер-ориентированной селекции по признакам питательной ценности. И наконец, обсуждаются генетические работы, посвященные весьма актуальной для селекции проблеме оптимального в тех или иных климатических условиях срока созревания плодов и их длительного хранения без потери качества.

Ключевые слова: томат; селекция; ДНК-маркер; патоген; устойчивость; срок созревания; лежкость.

Introduction

Tomato, *Solanum lycopersicum* L., is the second most important vegetable crop after cabbage. It belongs to the family Solanaceae, consisting of approximately 100 genera and

2500 species, including several plants of agronomic importance (potato, eggplant, pepper, tobacco). In 2012, due to the efforts of the International Consortium on sequencing the tomato genome, the genomes of the cultivar Heinz 1706 and

the wild ancestor of the tomato, *Solanum pimpinellifolium* L. were completely sequenced (DOI 10.1038/nature11119). The tomato ($2n = 2x = 24$) has a relatively compact genome of 950 MB. It contains about 35,000 genes and was subjected to two rounds of triploidization (120 million and 70 million years ago) in the course of evolution; the second round took place before the divergence of tomato and potato. It is believed that the process of polyploidization promoted the neofunctionalization of genes responsible for the ripening and chemical composition of fruits, leading to the formation of a fleshy fruit in tomato that is of great importance for the propagation of seeds (Howe, Smallwood, 1982). Sequencing data is available through the SOLGenomics Network (SGN) website (<http://solgenomics.net>). Tomato fruits are enriched with vitamins A and C, a number of minerals and other biologically active substances (BAS), including lycopene, which belongs to antioxidants (Rao A.V., Rao L.G., 2007).

The homeland of tomato is South America, where its wild and semi-cultural forms are still found. In the middle of the XVI century, the tomato came through Spain and Portugal to Europe and was first used as an ornamental plant, since the fruits of the tomato were considered inedible. At the end of the XVIII century, a tomato appeared in Russia and was also first cultivated for decorative purposes. The tomato became a vegetable crop thanks to the agronomist scientist A.T. Bolotov, who developed a seedling method of cultivation and a method of ripening (ripening of green fruits after their collection).

DNA markers

Currently, the presence of complete genomic sequences (see above) makes it possible to effectively search for various genes responsible for valuable traits, as well as corresponding DNA markers for marker-assisted selection (MAS) of new forms of tomato. A large number of these markers were developed, including: RFLP (restriction fragment length polymorphism) (Tanksley et al., 1992), as well as PCR markers, including RAPD (randomly amplified polymorphic DNA), AFLP (amplified length polymorphism fragments), SSR (simple repeating sequences) (Saliba-Colombani et al., 2000; Ohshima et al., 2009). To date, SNP (single nucleotide polymorphism) markers are the technology of choice, and within this technology methodological approaches have been successfully approved on tomato such as using of EST SNP analysis for high-performance genotyping (Shirasawa et al., 2010), wide-scale genomic sequencing to identify SNPs that affect protein functions (<http://plantl.kazusa.or.jp/tomato/>). Polymorphic markers for tomato genomic selection were developed based on DArT (DNA chip technology for studying diversity) (Van Schalkwyk et al., 2012).

However, it should be noted that, despite the many DNA markers developed, mainly markers for qualitative traits, such as specific resistance to pathogens, are currently used in practical breeding of tomato. As for quantitative traits (QTL), so far the use of appropriate markers is hindered by their weak linkage with these traits, low polymorphism, undesirable pleiotropic effects, and the lack of validation on diverse material of lines and varieties (Foolad, Panthee, 2012). In this regard, the problem of developing of new, effective molecular markers suitable for use on a wide range of varieties and populations remains actual.

Main directions of tomato breeding in Western Siberia

Tomato is a thermophilic culture and the climate of Western Siberia does not always favor to its productivity. In addition, the tomato is susceptible to numerous infectious diseases. This implies the need to obtain new varieties and hybrids capable of producing high yields and possessing a set of economically valuable traits, such as resistance to pathogens, ripening date corresponding to a short vegetation period, shelf life, etc. As known, MAS makes it possible to conduct selection for many traits simultaneously and allows significantly (2–3 times) to reduce the time of obtaining new varieties, compared with the classical breeding. However, no one variety or hybrid of tomato has been obtained in Siberia using MAS. In this regard, it seems relevant to summarize the main results obtained in the world on this culture with the help of MAS, focusing on those directions that correspond to the conditions of Western Siberia and adjacent territories.

Tomato resistance to pathogens

Most of the resistance genes were identified within wild-growing species and then, by crossing, were introduced in a cultivated tomato (Foolad, Panthee, 2012). In Siberia, fungal diseases of tomato are in the first place by importance, namely: late blight, leaf mould (in greenhouse), septoria blight (in field), fusarium wilt and verticillium wilt. Bacterial spot and bacterial canker are the most common bacterial diseases. Viral diseases are not so relevant for Siberia, although in some years epiphytotics occur.

Resistance to fungal diseases

Late blight caused by *Phytophthora infestans* oomycete, is one of the most devastating diseases of tomato in regions with high humidity and a cool climate, leading to yield loss up to 100 %. Losses can be in the form of a drop in yield, a lower quality of fruits, for example, a low specific weight, a decrease in shelf life, etc. Due to the large economic effect, the pathology and genetics of this disease have been intensively studied for many years. Three main resistance genes were identified in wild-growing tomato *S. pimpinellifolium*: *Ph-1*, *Ph-2* and *Ph-3*, which were mapped on chromosomes 7, 10 and 9, respectively (Black et al., 1996; Moreau et al., 1998). The strongest resistance gene, *Ph-3*, provides incomplete dominant resistance to a wide range of *P. infestans* isolates (Chunwongse et al., 2002). Analysis of its primary structure showed that it encodes a CC-NBS-LRR (coiled-coil nucleotide-binding leucine-rich repeat) – protein that belongs to the extensive NBS-LRR class of plant R-genes (Zhang et al., 2014). However, even this gene does not provide resistance to *Phytophthora* most aggressive isolates. In these cases, the most effective was the combination of two genes, *Ph-2* and *Ph-3*, which were successfully transferred to a number of commercial varieties using developed CAPS markers (Robbins et al., 2010; Zhang et al., 2014). Work on the isolation and analysis of new late blight resistance genes continues. In particular, a number of QTLs carrying resistance genes have been identified that have not yet been precisely localized (Merk, Foolad, 2012; Panthee et al., 2017).

Fusarium wilt. *Fusarium oxysporum* is a soil fungus that causes wilting disease in tomato. It affects all plant tissues and

can persist for a long time in the form of chlamydospores in the soil and plant residues, without losing virulence. Currently three races of this fungus were identified; in Russia, race 1 brings the most damage in greenhouses, race 2 occurs in some farms (Ignatova, 2001). Gene *I*, which provides high resistance to race 1, and gene *I-2*, which gives resistance to races 1 and 2, were mapped on the short and long arms of chromosome 11, respectively (Ori et al., 1997; Scott et al., 2004). These genes were most often used in breeding for resistance to *Fusarium*, however, recently, race 3 has become very common and the corresponding resistance gene has been mapped in detail on chromosome 7 (Lim et al., 2008). There are various linked PCR markers for each of the three genes; markers of resistance to races 1 and 3 are most effective (Barillas et al., 2008; Arens et al., 2010).

A kind of Fusarium wilt – Fusarium root rot, caused by another strain of *F. oxysporum*. Resistance was established in the induced mutant *S. peruvianum* and the only resistance gene *Fr 1* was mapped on chromosome 9 near the *Tm-2²* gene (Vakalounakis et al., 1997). Subsequently, RAPD markers for *Fr 1* (Tanyolac, Akkale, 2010) were developed, however, to date there are few commercial varieties and lines resistant to this disease.

Leaf mould is common in almost all the world and most often affects plants in greenhouse conditions. Affected leaves, flowers and young fruits turn yellow and then dry. The pathogenic agent is *Cladosporium fulvum*, a highly contagious, optional saprotroph. More than 20 major resistance genes have been identified and mapped on different chromosomes (Wang et al., 2007). In Russia, the most effective resistance genes *Cf-2*, *Cf-5*, *Cf-6*, *Cf-9* give resistance to races of the fungus 1, 3 and 4, however, due to the appearance of new races, at least two genes must be combined (Ignatova, 2001). Although a number of PCR markers have been associated with *Cf* genes (Grushetskaya et al., 2007; Wang et al., 2007; Truong et al., 2011), there is no data on their use in breeding.

Verticillium wilt is a widespread disease characterized by the following symptoms: wilting, discoloration and leaf fall, vascular tissues and root system necrosis. Verticillium wilt is caused by *Verticillium dahlia* and *V. albo-atrum*. In tomato, resistance to *Verticillium* is controlled by the *Ve* locus mapped on the short arm of chromosome 9 and consisting of two linked genes *Ve-1* and *Ve-2*, each of which provides resistance to certain pathogen races (Kawchuk et al., 2001; Fradin et al., 2009). PCR markers were obtained to discriminate tolerant and sensitive to *Verticillium* forms of tomato (Acciarri et al., 2007; Arens et al., 2010).

Resistance to bacterial pathogens

Bacterial cancer caused by the rod-shaped bacterium *Clavibacter michiganensis*, is a common tomato disease worldwide and one of the most difficult to control. Infection occurs through mechanically damaged tissues. Greenhouse tomatoes are most at risk. Mapping using crosses between *S. lycopersicum* and the resistant specimen *S. habrochaites* LA 407 allowed to identify and accurately map two large QTLs on chromosomes 2 (*Rcm2.0*) and 5 (*Rcm5.1*), which are responsible for 68 % of expressivity variation (Kabelka et al., 2002; Coaker, Francis, 2004). There are data on markers

(Coaker, Francis, 2004), however, there is no information on their use.

Bacterial spot is a common disease of tomato (especially in Western Siberia, Kazakhstan), which is caused by four species of rod-shaped bacteria *Xanthomonas* (races T1–T5). It is characterized by spotting of leaves, stems and fruits, accompanied by leaf fall, a decrease in the size of fruits and their immaturity, which leads to yield loss up to 100 %. Chemical control is not effective enough due to the development of resistance in the pathogen and multiple ways of its inoculation. Pathogen resistance has been found in a number of *S. lycopersicum* specimens, as well as in wild species, however, its use is greatly complicated by the diversity of pathogen races and the complex nature of resistance. In many cases, it is characterized by race specificity, but some genotypes exhibit multiple quantitative resistance, depending on external conditions. For example, the resistance of Hawaii 7998 *S. lycopersicum* line to race T1 ranges from reduced field symptoms to a hypersensitivity reaction (HR) in a greenhouse. This reaction is provided by three independent genes *Rx-1*, *Rx-2* (chromosome 1) and *Rx-3* (chromosome 5) (Wang et al., 1994; Yu et al., 1995). The participation of *Rx-3* locus was most reliably confirmed to which markers were developed, including CAPS marker L3-L1, which was used in breeding (Yang, Francis, 2005). The same line has strong HR-resistance to race T3 (both in the field and in the greenhouse), which is controlled by the *Rx-4* gene mapped on chromosome 11 (Wang et al., 2011).

Resistance to viruses

Tomato mosaic virus (ToMV) is one of the most stable viruses; crop losses when infected with ToMV reach 50 % or more. The disease is characterized by the appearance of a motley (mosaic) color of leaves, stems and fruits, followed by their deformation and fading. ToMV is highly contagious and is transmitted via mechanical contact, as well as insects: thrips, aphids, etc. In tomato three major resistance genes were revealed: *Tm-1*, *Tm-2* and *Tm-2²* (Ohmori et al., 1996; Sobir et al., 2000; Scott, 2007). The first gene, localized on chromosome 5, inhibits the synthesis of viral RNA by suppressing viral RNA replicase (Meshi et al., 1988). The *Tm-2* and *Tm-2²* genes, localized on chromosome 9, block the movement of the virus from cell to cell, and also cause HR (Meshi et al., 1989). The highest efficiency is observed when all three dominant genes are combined in the homo- or heterozygous state (Puchalsky, 2007). For each of these, PCR markers were developed (Dax et al., 1998; Sobir et al., 2000; Arens et al., 2010).

Tomato spotted wilt virus. The disease is caused by the tomato spotted wilt virus, TSWV. It leads to a decrease in crop yields (over 50 %) and deterioration in product quality. The TSWV virus has an extremely wide range of host plants, which creates a high risk of infection. Eight major resistance genes are known, including the dominant genes *Sw-1a*, *Sw-1b*, *Sw-5*, *Sw-6* and *Sw-7* and the recessive genes *sw-2*, *sw-3* and *sw-4* (Stevens et al., 1992). The most effective gene for resistance to TSWV, the *Sw-5* gene, is localized on the long arm of chromosome 9, and since it is race-specific, it is often used in practical breeding. However, there is a risk of overcoming *Sw-5* with new TSWV strains; virulence to this

resistance gene has been reported in several countries (Scott, 2007). A large number of PCR markers have been developed to detect *Sw-5* (Smiech et al., 2000; Langella et al., 2004; Garland et al., 2005).

Size and color of fruits, content in them biologically active substances

The trait of “uniform ripening” is determined by the genetic locus *uniform ripening* (*u*), which control the amount and distribution of chlorophyll in immature fruits (Bohn, Scott, 1945). The dominant allele *U* determines a normal, uneven maturation, in which the upper part of the immature fruit has a dark green and the lower – a light green color. Plants that are homozygous for the recessive *u* allele (*u/u*) produce uniformly ripening fruits that, in an immature state, have the same pale green color on all sides. The initial breeding led to the selection of such forms of tomato, because they are characterized by a uniform red color of ripe fruit. In 2012, localization of the *U* locus on the short arm of chromosome 10 was established using genetic mapping and the *GLK2* candidate gene was identified that encodes the Golden 2-like transcription factor, a regulator of chloroplast development (Powell et al., 2012). The authors sequenced this gene in varieties with *U/U* and *u/u* genotypes and found that in the first case, the *GLK2* gene encodes a complete regulatory protein of 310 amino acids in length, whereas in the case of the *u* allele, the synthesis of non-functional protein occurs due to premature stop codon which resulted from insertion of one nucleotide. Using genetic transformation, it was shown that this mutation blocking the *GLK2* gene is responsible for the uniform coloring phenotype and the associated decrease in the number of chloroplasts in fruits. The latter, in turn, leads to a decrease in the level of photosynthesis and a significant decrease in the content of soluble solids in the fruit juice. As a result, the cultural forms of tomato with the *u/u* genotype have lower taste and nutritional qualities, compared with the ancestral forms. In 2017, the Science published an article of D. Tieman et al. (2017), in which more than 300 modern and traditional tomato varieties were analyzed using genomic sequencing and chemical analysis. In this work, 28 compounds were identified that are responsible for the organoleptic qualities of tomato and then, based on the genome-wide analysis of associations (GWAS), a search was made for SNPs associated with the concentration of these chemical compounds. As a result, several major genes were identified that are responsible for the tomato flavor. Thus, the *Lin5* gene encodes an extracellular invertase that catalyzes the hydrolysis of sucrose to low molecular weight glucose and fructose. Alleles of this gene that are responsible for the alternative characteristics of modern and wild/old-fashioned varieties (low sugar content, large fruits vs. high content, small fruits) differ by only one SNP, leading to the substitution Asn→Asp. Another example, the *E8* gene, which regulates the synthesis of ethylene, hormone of maturation. In the overwhelming majority of modern varieties, this hormone has an increased activity, which leads to a higher concentration of methyl salicylate and guaiacol with an unpleasant smell, compared to the old varieties, while the “beneficial” aromatic substances are less concentrated. Three SNPs were identified in the regulatory regions of the *E8* gene, which appear to be responsible for

the indicated differences (Tieman et al., 2017).

The most important BAS of tomato fruits include carotenoids, a class of 40-carbon hydrocarbons, which are represented by orange, red and yellow pigments synthesized in various plant organs. These substances are involved in a variety of physiological processes of growth, development of plants, reactions to external stimuli. To date, the biosynthesis genes, as well as transcription factors and hormones that regulate the metabolism of carotenoids under the influence of external factors, have been established (Liu et al., 2015). In particular, key regulatory genes that determine the concentration of lycopene, the most common carotenoid-antioxidant of ripe tomatoes, have been identified. This substance is considered as an important biologically active component of the human diet, reducing the risk of cancer and cardiovascular diseases (Ford, Erdman, 2012). Recently, using genomic editing, the synthesis of lycopene in tomato fruits has been increased five times due to the knockout of genes responsible for the conversion of lycopene to β - and α -carotene (Li et al., 2018).

Specific polymorphisms that are responsible for particular varietal characteristics of the tomato fruit color were identified. The formation of a dark red color in the Black Cherry variety is caused by a mutation of the reading frame shift in the coding part of the lycopene- β -cyclase gene, leading to a loss of protein function. A similar mutation leading to a stop codon and shortened protein Psy 1 phytoene synthase underlies the yellow color of fruits (Aflitos et al., 2014).

The shape and size of the tomato fruit correlates with the number of seed chambers (locules). Two QTLs, *lc* and *fas*, have the maximum effect on these traits and can act synergistically, leading to an extremely high number of locules (Cong et al., 2008; Munos et al., 2011). *Fas* is the strongest gene (variation in the number of locules 2 more than 6), while *lc* acts weaker (3–4 locules). Two SNPs, T→C and A→G, are associated with the allele *lc^h* of a high number of locules. Analysis of the primary structure of the *lc* gene showed that all 2-chamber tomato varieties have the *lc^l* allele, and the 3, 4-chamber – allele *lc^h*. The *Fas* gene encodes a YABBY-like transcription factor (Cong et al., 2008). The *fas^h* allele appeared as a result of the inversion of the 294 kbp region on chromosome 11, that led to the shutdown of the *Fas* gene due to the spatial separation of exons 1 and 2 (Huang, van der Knaap, 2011).

Peculiarities of the formation of plants and fruit ripening

Determinancy. For greenhouse conditions tomato plants of an indeterminate type are most suitable. They are characterized by continuous growth and uniform ripening of fruits for several months. For field conditions of Siberia determinant genotypes are more acceptable, the main distinguishing feature of which is termination of shoot growth after the formation of 2–6 inflorescences. Such genotypes, as a rule, are early maturing, which prevents yield loss due to the short growing season.

Determinancy is controlled by the *SP* regulatory gene (*SELF PRUNING*), which controls the transition from the vegetative to the generative stage of development and is homologous to *FT* (*FLOWERING LOCUS T*) – gene of *Arabidopsis* (Pnueli et al., 1998). Determinant plants have the *sp/sp* genotype,

indeterminate – (*SP*–). There are at least six *SP* genes in the tomato genome. For one of them, *SP5G*, a mechanism of action was established that depends on photoperiod (Soyk et al., 2017). Like the *FT* gene, *SP5G* belongs to the flowering repressors. Under the influence of a long day, its expression is induced to a high level, which leads to suppression of flowering until the onset of a short day (indeterminant, wild phenotype). In a cultural tomato of determinant type, this effect of a long day on expression is reduced due to mutations in this gene. Using the CRISPR/Cas9 genomic editing, it was possible to obtain the null allele *SP5G* and thereby restore a determinant phenotype characterized by early flowering and increased productivity (Soyk et al., 2017).

Genes of slow ripening of fruits. Earlier, the pleiotropic genes responsible for the delayed fruit ripening period were revealed in tomato: *alcobaca* (*alc*), ripening inhibitor (*rin*) and non-ripening (*nor*) (Garg et al., 2008). In plants carrying these genes in a homozygous state, shelf life of fruits increased by 250–500 %; meanwhile they were less prone to the process of decay. However, such genotypes did not become widespread in commerce, due to the accompanying traits: pale coloring and poor taste. The fruits of heterozygous plants also had an increased shelf life (average between parental forms), resistance to decay, but at the same time they had acceptable color and taste for consumers. In addition, these plants had an increased yield, and such indicators as: the content of lycopene and dry matter, fruit consistency, ascorbic acid content were intermediate compared to their parents. As a result, the forms carrying the *alc*, *nor*, and *rin* genes are widely used in commercial tomato varieties in many countries (Garg et al., 2008).

In 2002 Science published an article devoted to the *rin* gene (Vrebalov et al., 2002). This gene is located on the short arm of chromosome 5 and encodes a MADS-box-transcription factor that regulates many different developmental genes, including those associated with ethylene biosynthesis. The *alc* and *nor* genes were also cloned and analyzed (Moore et al., 2002). The *alc* gene (synonym: *DFD*, delayed fruit deterioration) has several advantages for breeding, since it has a lower negative effect on fruit quality, color, aromatic properties and resistance to bacterial diseases (Garg et al., 2008). The *alc* recessive mutation is caused by a nonsynonymous T→A substitution at position 317 of the coding sequence, leading to the Val→Asp substitution (Casals et al., 2012). Using CRISPR/Cas9 in one of the varieties, the *ALC* allele was replaced by the *alc* allele by homologous recombination (Yu et al., 2017).

Functional male sterility

The low genetic diversity due to the mode of tomato reproduction (self-pollination) and the effect of the bottle neck during the introduction process make the successful breeding of tomato very difficult. The English scientist Ch. Rick first began to use the methods of introgression of genetic material from wild-growing to cultivated tomato (Rick, 1960) and most of the tomato varieties were obtained using hybridization.

In tomato, the production of hybrid seeds is laborious due to the need for isolation and castration of flowers, so the use of lines with the trait of functional male sterility (FMS) is the most effective way to obtain hybrid seeds. FMS is

caused by deviations in the development of the flower and in tomato includes the following types: *ex*, *ex-2*, *ps*, *ps-2* (Kuzemensky, 2004). The latter type is most widely used in tomato breeding. The stamens of plants of the *ps-2* type have the usual structure, fertile pollen grains, but the anthers are not opened. The *Ps-2* gene controlling this type of sterility was identified in chromosome 4, isolated and its primary structure was studied (Gorguet et al., 2009). It encodes the enzyme polygalacturonase, which affects the rigidity of the cell wall by digestion of pectins. The single mutation that disrupts splicing of mRNA, resulting in its aberrant forms is responsible for the *ps2* phenotype. A number of markers have been developed for the *Ps-2* gene: SNP (Gorguet et al., 2009), CAPS (Staniaszek et al., 2012), etc.

Conclusion

The work on the complete sequencing of the tomato genome and the construction of high-resolution genetic maps laid the foundation for a fast and effective search for genes responsible for important selection traits, as well as the development of DNA markers corresponding to these genes that can be used in marker-assisted selection of a new forms of tomato. Especially relevant for a temperate climate are markers of such traits as resistance to a number of common pathogens of various nature, valuable biologically active substances, for example, carotenoids, lycopene, sugars, etc., as well as gene markers that determine the optimal, early fruit ripening in conditions of short summer period and risk of autumn frosts. To date, key genes responsible for these traits have been identified and characterized, which makes it possible, on the basis of molecular markers, to develop strategies for crossing and selection for these genes, to perform their pyramiding, as well as targeted modification using modern genomic editing methods.

References

- Acciarri N., Rotino G.L., Tamiotti G., Valentino D., Voltattorni S., Sabatini E. Molecular markers for *Ve1* and *Ve2* *Verticillium* resistance genes from Italian tomato germplasm. *Plant Breed.* 2007;126:617-621. DOI 10.1111/j.1439-0523.2007.01398.x.
- Aflitos S., Schijlen E., de Jong H., de Ridder D., Smit S., Finkers R., Wang J., Zhang G., Li N., Mao L., Bakker F., Dirks R., Breit T., Gravendeel B., Huits H., Struss D., Swanson-Wagner R., van Leeuwen H., van Ham R.C., Fito L., Guignier L., Sevilla M., Ellul P., Ganko E., Kapur A., Reclus E., de Geus B., van de Geest H., Te Lintel Hekkert B., van Haarst J., Smits L., Koops A., Sanchez-Perez G., van Heusden A.W., Visser R., Quan Z., Min J., Liao L., Wang X., Wang G., Yue Z., Yang X., Xu N., Schranz E., Smets E., Vos R., Rauwerda J., Ursem R., Schuit C., Kerns M., van den Berg J., Vriezen W., Janssen A., Datema E., Jahrman T., Moquet F., Bonnet J., Peters S. Exploring genetic variation in the tomato (*Solanum* section *Lycopersicon*) clade by whole-genome sequencing. *Plant J.* 2014;80:136-148. DOI 10.1111/tpj.12616.
- Arens P., Mansilla C., Deinum D., Cavellini L., Moretti A., Roland S., van der Schoot H., Calvache D., Ponz F., Collonnier C., Mathis R., Smilde D., Caranta C., Vosman B. Development and evaluation of robust molecular markers linked to disease resistance in tomato for distinctness, uniformity and stability testing. *Theor. Appl. Genet.* 2010;120:655-664. DOI 10.1007/s00122-009-1183-2.

- Barillas A.C., Mejia L., Sanchez-Perez A., Maxwell D.P. CAPS and SCAR markers for detection of *I-3* gene introgression for resistance to *Fusarium oxysporium* f. sp. *lycopersici* race 3. Rpt. Tomato Genet. Coop. 2008;58:11-17.
- Black L.L., Wang T.C., Hanson P.M., Chen J.T. Late blight resistance in four wild tomato accessions: effectiveness in diverse locations and inheritance of resistance. *Phytopathology*. 1996;86:S24.
- Bohn G.W., Scott D.H. A second gene for uniform unripe fruit color in the tomato. *J. Hered.* 1945;36(6):169-172.
- Casals J., Pascual L., Cañizares J., Cebolla-Cornejo J., Casañas F., Nuez F. Genetic basis of long shelf life and variability into Penjar tomato. *Genet. Resour. Crop Evol.* 2012;59:219-229. DOI 10.1007/s10722-011-9677-6.
- Chunwongse J., Chunwongse C., Black L., Hanson P. Molecular mapping of the *Ph-3* gene for late blight resistance in tomato. *J. Horticult. Sci. Biotechnol.* 2002;77:281-286. DOI 10.1080/14620316.2002.11511493.
- Coaker G.L., Francis D.M. Mapping, genetic effects, and epistatic interaction of two bacterial canker resistance QTLs from *Lycopersicon hirsutum*. *Theor. Appl. Genet.* 2004;108:1047-1055. DOI 10.1007/s00122-003-1531-6.
- Cong B., Barrero L.S., Tanksley S.D. Regulatory change in YABBY-like transcription factor led to evolution of extreme fruit size during tomato domestication. *Nat. Genet.* 2008;40:800-804. DOI 10.1038/ng.144.
- Dax E., Livneh O., Aliskevicius E., Edelbaum O., Kedar N., Gavish N., Milo J., Geffen F., Blumenthal A., Rabinowich H.D., Sela I. A SCAR marker linked to the ToMV resistance gene, *Tm2(2)*, in tomato. *Euphytica*. 1998;101:73-77.
- Foolad M.R., Panthee D.R. Marker-assisted selection in tomato breeding. *Crit. Rev. Plant Sci.* 2012;31(2):93-123. DOI 10.1080/07352689.2011.616057.
- Ford N.A., Erdman J.W. Are lycopene metabolites metabolically active? *Acta Biochim. Pol.* 2012;59:1-4.
- Fradin E.F., Zhang Z., Juarez Ayala J.C., Castroverde C.D., Nazar R.N., Robb J., Liu C.M., Thomma B.P. Genetic dissection of verticillium wilt resistance mediated by tomato *Ve1*. *Plant Physiol.* 2009;150: 320-332. DOI 10.1104/pp.109.136762.
- Garg N., Cheema D.S., Pathak D. Heterosis breeding in tomato involving *rin*, *nor* and *alc* alleles: A review of literature. *Adv. Hort. Sci.* 2008;22(1):54-62.
- Garland S., Sharman M., Persley D., McGrath D. The development of an improved PCR-based marker system for *Sw-5*, an important TSWV resistance gene of tomato. *Aust. J. Agric. Res.* 2005;56: 285-289.
- Gorguet B., Schipper D., van Lammeren A., Visser R.G.F., van Heusden A.W. *ps-2*, the gene responsible for functional sterility in tomato, due to non-dehiscent anthers, is the result of a mutation in a novel polygalacturonase gene. *Theor. Appl. Genet.* 2009;118:1199-1209. DOI 10.1007/s00122-009-0974-9.
- Grushetskaya Z.E., Lemesh V.A., Poliksenova V.D., Khotyleva L.V. Mapping of the *Cf-6* tomato leaf mould resistance locus using SSR markers. *Russ. J. Genet.* 2007;43:1266-1270. DOI 10.1134/S1022795407110099.
- Howe H.F., Smallwood J. Ecology of seed dispersal. *Annu. Rev. Ecol. Syst.* 1982;13:201-228.
- Huang Z., van der Knaap E. Tomato fruit weight 11.3 maps close to fasciated on the bottom of chromosome 11. *Theor. Appl. Genet.* 2011; 123(3):465-474. DOI 10.1007/s00122-011-1599-3.
- Ignatova S.I. The role of tomato hereditary potential for resistance in the system of integrated protection in protected ground. *Gavrish.* 2001;6:18-20. (in Russian)
- Kabelka E., Franchino B., Francis D.M. Two loci from *Lycopersicon hirsutum* LA407 confer resistance to strains of *Clavibacter michiganensis* subsp. *michiganensis*. *Phytopathology*. 2002;92:504-510. DOI 10.1094/PHYTO.2002.92.5.504.
- Kawchuk L.M., Lynch D.R., Hachey J., Bains P.S., Kulcsar F. Identification of a codominant amplified polymorphic DNA marker linked to the verticillium wilt resistance gene in tomato. *Theor. Appl. Genet.* 1994;89:661-664.
- Kuzemsky A.V. Studies of Mutant Forms of Tomato with Regard to Breeding. Kharkov, 2004. (in Russian)
- Langella R., Ercolano M.R., Monti L.M., Frusciante L., Barone A. Molecular marker assisted transfer of resistance to TSWV in tomato elite lines. *J. Horticult. Sci. Biotechnol.* 2004;79:806-810. DOI 10.1080/14620316.2004.11511846.
- Li X., Wang Y., Chen S., Tian H., Fu D., Zhu B., Luo Y., Zhu H. Lycopene is enriched in tomato fruit by CRISPR/Cas9-mediated multiplex genome editing. *Front. Plant Sci.* 2018;9:1-12. DOI 10.3389/fpls.2018.00559.
- Lim G., Wang G.P., Hemming M., McGrath D.J., Jones D.A. High resolution genetic and physical mapping of the I-3 region of tomato chromosome 7 reveals almost continuous microsynteny with grape chromosome 12 but interspersed microsynteny with duplications on *Arabidopsis* chromosomes 1, 2 and 3. *Theor. Appl. Genet.* 2008; 118:57-75. DOI 10.1007/s00122-008-0876-2.
- Liu L., Shao S.Z., Zhang Z.M. Regulation of carotenoid metabolism in tomato. *Mol. Plant.* 2015;8:28-39. DOI 10.1016/j.molp.2014.11.006.
- Merk H.L., Foolad M.R. Parent-offspring correlation estimate of heritability for late blight resistance conferred by an accession of the tomato wild species *Solanum pimpinellifolium*. *Plant Breed.* 2012; 131:203-210.
- Meshi T., Motoyoshi F., Adachi A., Watanabe Y., Takamatsu N., Okada Y. Two concomitant base substitutions in the putative replicase genes of tobacco mosaic virus confer the ability to overcome the effects of a tomato resistance gene, *Tin-1*. *EMBO J.* 1988;7:1575-1581.
- Meshi T., Motoyoshi F., Maeda T., Yoshiwoka S., Watanabe H., Okada Y. Mutations in the tobacco mosaic virus 30-kDa protein gene overcome *Tm-2* resistance in tomato. *Plant Cell.* 1989;1:515-522.
- Moore S., Vrebalov J., Payton P., Giovannoni J. Use of genomics tools to isolate key ripening genes and analyse fruit maturation in tomato. *J. Exp. Bot.* 2002;53:2023-2030. DOI 10.1093/jxb/erf057.
- Moreau P., Thoquet P., Olivier J., Laterrot H., Grimsley N. Genetic mapping of *Ph-2*, a single locus controlling partial resistance to *Phytophthora infestans* in tomato. *Mol. Plant-Microbe Interact.* 1998; 11:259-269. DOI 10.1094/MPMI.1998.11.4.259.
- Muñoz S., Ranc N., Botton E., Bérard A., Rolland S., Duffé P., Carretero Y., Le Paslier M.-C., Delalande C., Bouzayen M., Brunel D., Causse M. Increase in tomato locule number is controlled by two single nucleotide polymorphisms located near *WUSCHEL*. *Plant Physiol.* 2011;156:2244-2254. DOI 10.1104/pp.111.173997.
- Ohmori T., Murata M., Motoyoshi F. Molecular characterization of RAPD and SCAR markers linked to the *Tm-1* locus in tomato. *Theor. Appl. Genet.* 1996;92:151-156. DOI 10.1007/BF00223369.
- Ohyama A., Asamizu E., Negoro S., Miyatake K., Yamaguchi H., Tabata S., Fukuoka H. Characterization of tomato SSR markers developed using BAC-end and cDNA sequences from genome databases. *Mol. Breed.* 2009;23(4):685-691. DOI 10.1007/s11032-009-9265-z.

- Ori N., Eshed Y., Paran I., Presting G., Aviv D., Tanksley S., Zamir D., Fluhr R. The *I2C* family from the wilt disease resistance locus I2 belongs to the nucleotide binding, leucine-rich repeat superfamily of plant resistance genes. *Plant Cell*. 1997;9:521-532. DOI 10.1105/tpc.9.4.521.
- Panthee D.R., Piotrowski A., Ibrahim R. Mapping quantitative trait loci (QTL) for resistance to late blight in tomato. *Int. J. Mol. Sci.* 2017;18:1589. DOI 10.3390/ijms18071589.
- Pnueli L., Carmel-Goren L., Hareven D., Gutfinger T., Alvarez J., Ganai M., Zamir D., Lifschitz E. The *SELF-PRUNING* gene of tomato regulates vegetative to reproductive switching of sympodial meristems and is the ortholog of *CEN* and *TFL1*. *Development*. 1998; 125:1979-1989.
- Powell A.L.T., Nguyen C.V., Hill T., Cheng K.L., Figueroa-Balderas R., Aktas H., Ashrafi H., Pons C., Fernández-Muñoz R., Vicente A., Lopez-Baltazar J., Barry C.S., Liu Y., Chetelat R., Granell A., Van Deynze A., Giovannoni J., Bennett A.B. Uniform ripening encodes a Golden 2-like transcription factor regulating tomato fruit chloroplast development. *Science*. 2012;336:1711-1715. DOI 10.1126/science.1222218.
- Pukhalskij V.A., Odintsova T.I., Izvekova L.I., Andreeva E.N., Korostyleva T.I., Istomina E.A., Slavokhotova A.A., Shiyun A.N., Kozlovskaya G.V., Obolenkova L.A., Badaeva E.D., Bilinskaya E.N. The problems of natural and induced immunity in plants. To the development of ideas N.I. Vavilova. *Informatsionny Vestnik VOGIS = The Herald of Vavilov Society for Geneticists and Breeders*. 2007;11(3/4):631-649. (in Russian)
- Rao A.V., Rao L.G. Carotenoids and human health. *Pharmacol. Res.* 2007;55:207-216. DOI 10.1016/j.phrs.2007.01.012.
- Rick C.M. Hybridization between *Lycopersicon esculentum* and *Solanum pennellii*: phylogenetic and cytogenetic significance. *Proc. Natl. Acad. Sci. USA*. 1960;46:78-82.
- Robbins M.D., Masud M.A.T., Panthee D.R., Gardner R.G., Francis D., Stevens M.R. Marker-assisted selection for coupling phase resistance to tomato spotted wilt virus and *Phytophthora infestans* (late blight) in tomato. *HortScience*. 2010;45:1424-1428.
- Saliba-Colombani V., Causse M., Gervais L., Philouze J. Efficiency of RFLP, RAPD, and AFLP markers for the construction of an intraspecific map of the tomato genome. *Genome*. 2000;43:29-40. DOI 10.1139/g99-096.
- Scott J.W. Breeding for resistance to viral pathogens. Eds. M.K. Razdan, A.K. Mattoo. *Genetic Improvement of Solanaceous Crops. Vol. 2. Tomato*. Enfield: Science Publishers, 2007;457-485.
- Scott J.W., Agrama H.A., Jones J.P. RFLP-based analysis of recombination among resistance genes to *Fusarium* wilt races 1, 2, and 3 in tomato. *J. Am. Soc. Hort. Sci.* 2004;129:394-400.
- Shirasawa K., Isobe S., Hirakawa H., Nakamura Y., Sato S., Tabata S. SNP discovery and linkage map construction in cultivated tomato. *DNA Res.* 2010;17(6):381-391. DOI 10.1093/dnares/dsq024.
- Smiech M., Rusinowski Z., Malepszy S., Niemirowicz-Szczytt K. New RAPD markers of tomato spotted wilt virus (TSWV) resistance in *Lycopersicon esculentum* Mill. *Acta Physiol. Plantarum*. 2000;22: 299-303.
- Sobir O.T., Murata M., Motoyoshi F. Molecular characterization of the SCAR markers tightly linked to the *Tm-2* locus of the genus *Lycopersicon*. *Theor. Appl. Genet.* 2000;101:64-69.
- Soyk S., Müller N.A., Park S.J., Schmalenbach I., Jiang K., Hayama R., Zhang L., Van Eck J., Jiménez-Gómez J.M., Lippman Z.B. Variation in the flowering gene *SELF PRUNING 5G* promotes day-neutrality and early yield in tomato. *Nat. Genet.* 2017;49:162-168. DOI 10.1038/ng.3733.
- Staniaszek M., Szajko K., Kozik E.U., Nowakowska M., Marczewski W. The novel *ps* and *ps-2* specific markers for selection of functional male sterile tomato lines in breeding programs and hybrids seed production. *J. Agr. Sci.* 2012;4(10):61-67. DOI 10.5539/jas.v4n10p61.
- Stevens M.R., Scott S.J., Gergerich R.C. Inheritance of gene for resistance to tomato spotted wilt virus (TSWV) from *Lycopersicon peruvianum* Mill. *Euphytica*. 1992;59:9-17. DOI 10.1007/BF00025356.
- Tanksley S.D., Ganai M.W., Prince J.P., de Vicente M.C., Bonierbale M.W., Broun P., Fulton T.M., Giovannoni J.J., Grandillo S., Martin G.B., Messeguer R., Miller J.C., Miller L., Paterson A.H., Pineda O., Riider M.S., Wing R.A., Wu W., Young N.D. High-density molecular linkage maps of the tomato and potato genomes. *Genetics*. 1992;132:1141-1160.
- Tanyolac B., Akkale C. Screening of resistance genes to fusarium root rot and fusarium wilt diseases in F3 family lines of tomato (*Lycopersicon esculentum*) using RAPD and CAPS markers. *Afr. J. Biotech.* 2010;9:2727-2730.
- Tieman D., Zhu G., Resende M.F.R., Lin T., Nguyen C., Bies D., Rambla J.L., Beltran K.S.O., Taylor M., Zhang B., Ikeda H., Liu Z., Fisher J., Zemach I., Monforte A., Zamir D., Granell A., Kirst M., Huang S., Klee H. A chemical genetic roadmap to improved tomato flavor. *Science*. 2017;355(6323):391-394. DOI 10.1126/science.aal1556.
- Truong H.T.H., Choi H.-S., Cho M.C., Lee H.E., Kim J.H. Use of *Cf-9* gene-based markers in marker-assisted selection to screen tomato cultivars with resistance to *Cladosporium fulvum*. *Hort. Environ. Biotechnol.* 2011;52:204-210. DOI 10.1007/s13580-011-0164-y.
- Vakalounakis D.J., Laterrot H., Moretti A., Ligoixigakis E.K., Smardas K. Linkage between *Frl* (*Fusarium oxysporum* f. sp. radialis-lycopersici resistance) and *Tm-2* (tobacco mosaic virus resistance-2) loci in tomato (*Lycopersicon esculentum*). *Ann. Appl. Biol.* 1997; 130:319-323. DOI 10.1111/j.1744-7348.1997.tb06835.x.
- Van Schalkwyk A., Wenzl P., Smit S., Lopez-Cobollo R., Kilian A., Bishop G., Hefer C., Berger D.K. Bin mapping of tomato diversity array (DARt) markers to genomic regions of *Solanum lycopersicum* × *Solanum pennellii* introgression lines. *Theor. Appl. Genet.* 2012;124:947-956. DOI 10.1007/s00122-011-1759-5.
- Vrebalov J., Ruezinsky D., Padmanabhan V., White R., Medrano D., Drake R., Schuch W., Giovannoni J. MADS-box gene necessary for fruit ripening at the tomato ripening-inhibitor (*Rin*) locus. *Science*. 2002;296:343-346. DOI 10.1126/science.1068181.
- Wang A.X., Meng F.J., Xu X.Y. Development of molecular markers linked to *Cladosporium fulvum* resistant gene *Cf-6* in tomato by RAPD and SSR methods. *HortScience*. 2007;42:11-15.
- Wang H., Hutton S.F., Robbins M.D., Sim S.-C., Scott J.W., Yang W., Jones J.B., Francis D.M. Molecular mapping of hypersensitive resistance to race T3 of tomato bacterial spot from Hawaii 7981 maps to chromosome 11. *Phytopathology*. 2011;101:1217-1223. DOI 10.1094/PHYTO-12-10-0345.
- Wang J.F., Jones J.B., Scott J.W., Stall R.E. Several genes in *Lycopersicon esculentum* control hypersensitivity to *Xanthomonas campestris* pv. *vesicatoria*. *Phytopathology*. 1994;84:702-706. DOI 10.1094/Phyto-84-702.
- Yang W., Francis D.M. Marker-assisted selection for combining resistance to bacterial spot and bacterial speck in tomato. *J. Am. Soc. Hort. Sci.* 2005;130:716-721. DOI 10.21273/JASHS.130.5.716.

- Yu Q.-H., Wang B., Li N., Tang Y., Yang S., Yang T., Xu J., Guo C., Yan P., Wang Q., Asmutola P. CRISPR/Cas9-induced targeted mutagenesis and gene replacement to generate long shelf life tomato lines. *Sci. Rep.* 2017;7:11874. DOI 10.1038/s41598-017-12262-1.
- Yu Z.H., Wang J.F., Stall R.E., Vallejos C.E. Genomic localization of tomato genes that control a hypersensitive reaction to *Xanthomonas campestris* pv. *vesicatoria* (Doidge) Dye. *Genetics*. 1995;141:675-682.
- Zhang C., Liu L., Wang X., Vossen J., Li G., Li T., Zheng Z., Gao J., Guo Y., Visser R.G.F., Li J., Bai Y., Du Y. The *Ph-3* gene from *Solanum pimpinellifolium* encodes CC-NBS-LRR protein conferring resistance to *Phytophthora infestans*. *Theor. Appl. Genet.* 2014;127:1353-1364. DOI 10.1007/s00122-014-2303-1.

ORCID ID

A.B. Shcherban orcid.org/0000-0003-1000-8228

Acknowledgements. This work was supported by the project of the Ministry of Education and Science of the Russian Federation No. 0324-2019-0039.

Conflict of interest. The authors declare no conflict of interest.

Received March 25, 2019. Revised May 17, 2019. Accepted May 20, 2019.