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Study of the influence of introgression from chromosome 2 of the A_t subgenome of cotton *Gossypium barbadense* L. during backcrossing with the original lines of *G. hirsutum* L.

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Abstract. The creation of chromosome substitution lines containing one pair of chromosomes from a related species is one method for introgression of alien genetic material. The frequency of substitutions in different chromosomes of the genome varies due to the selective transmission of alien chromosomes through the gametes of hybrids. The use of monosomic lines with identified univalent chromosomes and molecular genetic SSR markers at the seedling stage allowed rapid screening of the identity of the alien chromosome in backcross hybrids, significantly accelerating and facilitating the backcrossing process for the creation of new chromosome substitution cotton lines. As a result of studying the process of transmission of chromosome 2 of the A_t subgenome of the cotton plant *G. barbadense* L. during backcrossing of four original monosomic lines of *G. hirsutum* L. with monosomic backcross hybrids with substitution of chromosome 2 of the A_t subgenome, the following specific consequences of the introgression of this chromosome were revealed: decreased crossability, setting and germination of hybrid seeds; differences in the frequency and nature of transmission of chromosome 2 of the A_t subgenome of the cotton plant *G. barbadense*; regularity of chromosome behavior in meiosis; a high meiotic index; a significant decrease in pollen fertility in backcross monosomic hybrids BC₁F₁; specific morphobiological characteristics of monosomic backcrossed plants, such as delayed development of vegetative and generative organs; dwarfism; reduced foliage; and poor budding and flowering during the first year of vegetation. All of these factors negatively impact the study and backcrossing of monosomic hybrids and significantly complicate and delay the creation of chromosome-substituted forms concerning chromosome 2 of the A_t subgenome of cotton, *G. barbadense*. These specific changes likely occurred as a result of hybrid genome reorganization and introgression of alien chromatin. Furthermore, the effectiveness of using molecular genetic microsatellite (SSR) markers to monitor backcrossing processes and eliminate genetic material from the Pima 3-79 donor line of *G. barbadense* for the selection of genotypes with alien chromosome substitutions has been demonstrated.

Key words: cotton; *G. hirsutum*; *G. barbadense*; monosomic lines; chromosome-substituted hybrids; backcrossing; SSR markers

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Изучение влияния интровергессии хромосомы 2 A_t-субгенома хлопчатника вида *Gossypium barbadense* L. при беккроссировании исходными линиями вида *G. hirsutum* L.

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Аннотация. Создание хромосомно-замещенных линий, содержащих одну пару хромосом родственного вида, является одним из способов интровергессии чужеродного генетического материала. Известно, что частота встречаемости замещений по разным хромосомам генома различается по причине избирательности процесса трансмиссии чужеродных хромосом через гаметы гибридов. Использование моносомных линий с

идентифицированными унивалентными хромосомами и молекулярно-генетических SSR-маркеров на стадии проростков позволило осуществить быстрый скрининг идентичности чужеродной хромосомы у беккроссовых гибридов, что значительно ускорило и облегчило процесс беккроссирования при создании новых хромосомно-замещенных линий хлопчатника. В ходе изучения процесса передачи хромосомы 2 A_t -субгенома хлопчатника *Gossypium barbadense* L. при беккроссировании четырех исходных моносомных линий вида *G. hirsutum* L. моносомными беккроссовыми гибридами с замещением хромосомы 2 A_t -субгенома были выявлены: снижение скрещиваемости, завязываемости и всхожести гибридных семян; отличия в частоте и характере передачи хромосомы 2 A_t -субгенома хлопчатника *G. barbadense*; регулярность поведения хромосом в мейозе и высокий мейотический индекс; значительное снижение фертильности пыльцы у беккроссовых моносомных гибридов BC_1F_1 ; специфические морфобиологические особенности моносомных беккроссовых растений в виде задержки развития вегетативных и генеративных органов, низкорослости и сниженной облиственности, а также слабой бутонизации и цветения в течение первого года вегетации. По-видимому, такие специфические изменения происходили вследствие реорганизации гибридного генома и интродукции чужеродного хроматина. Кроме того, была продемонстрирована эффективность использования молекулярно-генетических микросателлитных (SSR) маркеров для контролирования процессов беккроссирования и элиминации генетического материала донорной линии Pima 3-79 вида *G. barbadense* с целью отбора генотипов с чужеродным замещением хромосом.

Ключевые слова: хлопчатник; *G. hirsutum*; *G. barbadense*; моносомные линии; хромосомно-замещенные гибриды; беккроссирование; SSR-маркеры

Introduction

Currently, the lack of genetic diversity in the cultivated cotton plant *Gossypium hirsutum* L. ($2n = 52$, AD_1) hinders the development of breeding programs (Wendel, 1989). One method of introducing alien genetic material is the creation of chromosome substitution lines containing one pair of chromosomes from a related species, which substitutes the homeologous pair of chromosomes because the substitution occurs in strict accordance with chromosome homeology (Shchapova, Kravtsova, 1990).

Lines with alien chromosome substitutions were created in the USA via three tetraploid cotton species (*G. barbadense* L., *G. tomentosum* Nutt. ex Seem and *G. mustelinum* Miers ex Watt) (Saha et al., 2004, 2006, 2013), where the largest number of lines were obtained from the *G. barbadense* species, and a study was conducted on the effects of substitution on valuable fibre quality traits (Saha et al., 2004, 2010, 2020; Jenkins et al., 2006, 2007). However, the active use of these lines by other researchers indicated the absence of introgression of the entire chromosome or a chromosomal region in some of these lines (Gutiérrez et al., 2009; Saha et al., 2015; Ulloa et al., 2016; Fang et al., 2023), so only 13 CS-B (chromosome substitution) lines out of 20 had “significant introgression” from the *G. barbadense* species. However, the reasons for the lack of substitution of alien chromosomes in the lines of the American collection have never been clarified.

Earlier, at the National University of Uzbekistan named after Mirzo Ulugbek a collection of new monosomic lines of *G. hirsutum* cotton was created via various irradiation methods. Univalent chromosomes of 35 of these lines, which are deficient in individual chromosomes, were identified via translocation and molecular genetic SSR markers (Sanamyan et al., 2014, 2022). Four monosomic lines were deficient in chromosome 2, 18 lines were deficient in chromosome 4, five lines were deficient in chromosome 6, one line was deficient in chromosome 7, and another line was deficient in chromosome 12 of the A_t subgenome of cotton. There is also one monosomic line on chromosomes 17, 18, 21 and 22 of the D_t subgenome of cotton and two telocentric lines on chromosomes 6 and 11 of the A_t subgenome (Sanamyan et al., 2016a, b; Sanamyan, Bobokhujaev, 2019).

To create chromosome substitution cotton lines, we developed a new scheme based on cytogenetic and molecular genetic methods (Sanamyan et al., 2022). The use of monosomic lines with identified univalent chromosomes and molecular genetic SSR markers at the seedling stage allowed rapid screening of the identity of alien chromosomes in backcross hybrids, significantly accelerating and facilitating the creation of chromosome substitution lines.

Experiments involving many hybrids across multiple generations have revealed new effects of alien chromosome transmission in backcrossed progeny. The aim of our study was to analyse the transmission patterns of chromosome 2 of the A_t subgenome of cotton during backcrossing of four original monosomic lines of *G. hirsutum* with the donor line Pima 3-79 of *G. barbadense* to create *G. hirsutum/G. barbadense* lines with alien chromosome substitutions. We assessed the crossability, seed set, and germination of hybrid seeds, estimated the frequency and pattern of chromosome 2 transmission, studied chromosome behavior during meiosis, and identified specific morphobiological features of backcrossed monosomic plants in the first year of vegetative growth.

Materials and methods

Plant material. The cotton cytogenetic collection of the National University of Uzbekistan named after M. Ulugbek is characterized by the presence of four monosomic lines (Mo11, Mo16, Mo19, and Mo93) deficient in chromosome 2 of the A_t subgenome of the tetraploid cotton species *G. hirsutum* ($2n = 52$, AD_1) (Sanamyan, Bobokhujaev, 2019).

The Pima 3-79 line, which was obtained from a doubled haploid and is the genetic standard for this cotton species in the United States (Endrizzi et al., 1985), was used as the donor parent of the substitution chromosome (CS) from the *G. barbadense* species.

Backcross hybrids obtained from crossing four monosomic lines (Mo11, Mo16, Mo19, and Mo93) deficient in chromosome 2 of the A_t subgenome of the *G. hirsutum* species with previously obtained monosomic F_1 hybrids, which had monosomy for the same chromosomes as the original monosomic plants (Sanamyan et al., 2016b), were studied. All plants of the original monosomic lines and backcross hybrids were

maintained year-round in a film greenhouse at the National University of Uzbekistan.

Cytological analyses, as well as DNA extraction and genotyping, were performed according to methods described previously (Sanamyan et al., 2023). Elimination of *G. barbadense* chromosomes in the monosomic cotton hybrid BC₂F₁ was determined by the absence of marker amplification on the *G. barbadense* chromosomes (paternal) and the presence of only allele-specific PCR products of *G. hirsutum* (maternal).

Results

Crossability, seed set, and germination of BC₁F₁ hybrid seeds involving monosomic lines deficient in chromosome 2 of the A_t subgenome of *G. hirsutum*. Four monosomic lines of *G. hirsutum* deficient in chromosome 2 of the A_t subgenome were crossed with F₁ hybrids (Mo×Pima 3-79), which were monosomic for the same chromosomes as the original monosomic line. All four BC₁F₁ variants were characterized by a significant decrease in the percentage of crossability (from 33.33 to 9.09 %) (Supplementary Material 1)¹ compared with F₁ hybrids (from 68.75 to 50.00 %) (Sanamyan et al., 2022).

The hybrid seed set of the BC₁F₁ plants also decreased (from 52.94±12.11 to 37.31±5.91 %) compared with that of the F₁ hybrids (from 57.69±9.69 to 31.33±5.09 %). Compared with that of the F₁ hybrids, the germination of the backcrossed BC₁F₁ seeds decreased from 88.89 to 51.61 % (Supplementary Material 1) (from 100 to 71.43 %).

Identification of chromosome 2 substitutions in the A_t subgenome of *G. barbadense* in BC₁F₁ hybrids via chromosome-specific molecular genetic markers. For molecular analysis of backcrossed BC₁F₁ plants, the principles of deletion molecular analysis were used (Liu et al., 2000; Gutiérrez et al., 2009). The study was conducted according to our previously proposed scheme for producing chromosome substitution cotton lines (Sanamyan et al., 2022). Molecular genetic analysis of the backcrossed forms was performed on backcrossed cotton seedlings at the 3–5 true leaf stage before they were transplanted into greenhouse soil to accelerate the isolation of monosomic chromosomes from the donor species. Among the plants harboring all the BC₁F₁ variants, those with genomes containing an alien chromosome 2 of the A_t subgenome of the *G. barbadense* species were identified.

Previously, among the BC₁F₁ hybrids in the BC₁F₁(Mo11×F₁(766₃)) cross variant in four backcross families (9_n, 10_n, 78_n, and 79_n), no hybrid seedlings with polymorphic alleles from the *G. barbadense* species were found, which indicated the absence of substitutions in these monosomes. Later, in a similar variant of the experiment in the backcross family (494_n), one (494₂) monosomic backcross seedling was identified, which was characterized by the presence of polymorphic alleles only from *G. barbadense*, whereas the alleles of the L-458 line of the *G. hirsutum* species were not detected on the basis of the localization of chromosome-specific SSR markers, BNL834, BNL1434, BNL1897, BNL3590, BNL3971, BNL3972, CIR381, and JESPR179. Since all the above-mentioned markers were previously localized on chromosome 2 of the A_t subgenome of cotton (Liu et al., 2000; Gutiérrez et al., 2009; Yu et al., 2011; Saha et al., 2015; Wang et al., 2016), the

obtained data indicated the presence of substitutions on this chromosome (Supplementary Materials 2, 3, 19).

Previously, in the BC₁F₁(Mo16×F₁(98₆)) variant, four seedlings (922₂, 922₈, 923₇, 923₈) with a substitution on chromosome 2 of the *G. barbadense* species were already found in two hybrid backcross families (922_n and 923_n) (Sanamyan et al., 2022); however, the elimination of the alien chromosome in BC₂F₁ required re-examination of this variant, where three backcross seedlings (495₅, 497₁ and 497₄) were found in two new families, which had only alleles from *G. barbadense*, whereas the alleles of *G. hirsutum* were absent, which indicated the localization of seven chromosome-specific SSR markers: BNL834, TMB0471, JESPR101, JESPR179, CIR376, DPL0674, and NAU2277, previously localized on chromosome 2 of the A_t subgenome (Gutiérrez et al., 2009; Yu et al., 2011; Saha et al., 2015; Wang et al., 2016) and confirmed the presence of a substitution on this chromosome (Supplementary Materials 4–6, 19).

SSR-based deletion analysis in the BC₁F₁(Mo19×F₁(769₄)) and BC₁F₁(Mo93×F₁(516₄)) combinations with a putative substitution of chromosome 2 of the A_t subgenome of cotton allowed us to detect alleles of the *G. barbadense* species in one backcross seedling (8₈) in the first variant and nine seedlings (7₁, 7₂, 7₃, 88₁, 88₄, 88₇, 89₃, 89₄ and 89₆) in the second, whereas alleles of the *G. hirsutum* species were absent. Since chromosome-specific SSR markers BNL3545, BNL3971, JESPR101, JESPR179, CIR376, and DPL0674 were previously localized on chromosome 2 of the A_t subgenome of cotton (Liu et al., 2000; Gutiérrez et al., 2009; Yu et al., 2011; Saha et al., 2015; Wang et al., 2016), the obtained data indicated the presence of substitution of this chromosome in the studied seedlings (Fig. 1, Supplementary Materials 7–9, 19).

A study of meiosis in BC₁F₁ hybrids with identified univalents. Chromosome pairing at the metaphase I (MI) stage of meiosis was studied in 17 monosomic plants in four backcross variants obtained from crosses of monosomic lines of the *G. hirsutum* species with interspecific monosomic F₁ hybrids (Mo×Pima 3-79). Seven monosomics were found among the backcrossed plants in the BC₁F₁(Mo16×F₁(98₆)) variant, one each in the BC₁F₁(Mo11×F₁(766₃)) and BC₁F₁(Mo19×F₁(769₄)) variants, and eight in the BC₁F₁(Mo93×F₁(516₄)) variant.

Analysis of meiotic MI in BC₁F₁ monosomic plants, where all backcrossed monosomic plants had univalent *G. barbadense* chromosomes, revealed a modal chromosome pairing with 25 bivalents and one univalent chromosome, characteristic of tetraploid monosomic cotton plants (Supplementary Material 10). Analysis of the size of univalents in monosomic BC₁F₁ confirmed the large size of chromosome 2 of *G. barbadense* in all four crossing variants (Fig. 2), which indicated that this chromosome belongs to the A_t subgenome of cotton.

Most of the studied BC₁F₁ monosomics were characterized by a high meiotic index (up to 95.74±0.47) and a small number of tetrads with micronuclei (up to 3.86±0.65 %), with the exception of two monosomics (922₈ and 7₃) with a reduced meiotic index (89.04±0.94 and 88.57±1.13, respectively) and an increased number of tetrads with micronuclei (up to 4.75±0.64 and 5.15±0.78 %, respectively) (Fig. 3, Supplementary Material 11).

¹ Supplementary Materials 1–20 are available at:
<https://vavilovj-icg.ru/download/pict-2025-29/appx45.pdf>

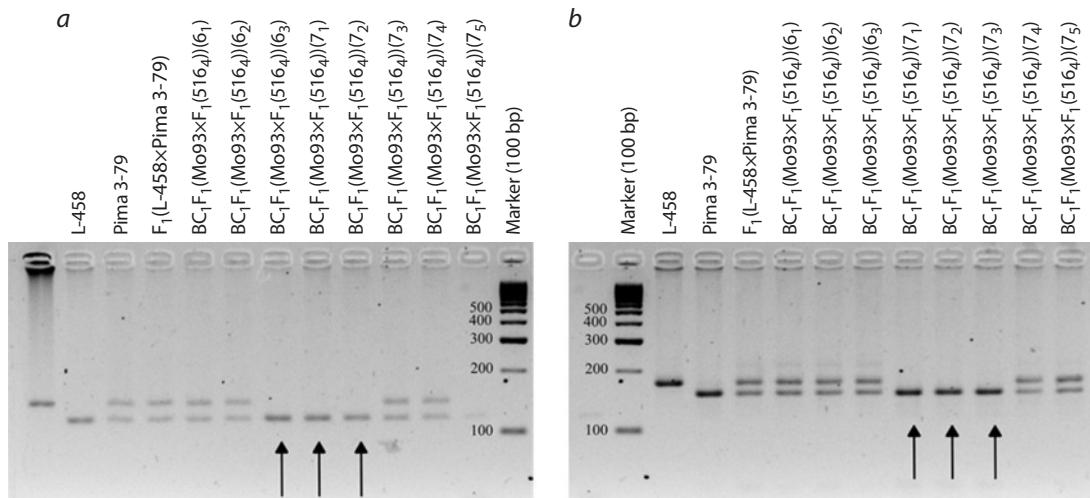


Fig. 1. Electropherogram of DNA amplicons of SSR markers in hybrid seedlings BC₁F₁(Mo93×F₁(516₄)) on chromosome 2 of the A_t subgenome of cotton.

a – BNL3971; b – JESPR179.

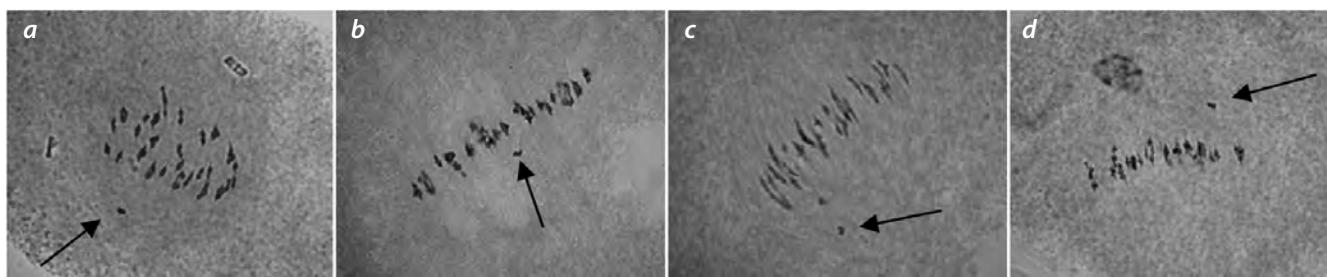


Fig. 2. Chromosome configurations in metaphase I of meiosis in BC₁F₁ hybrid plants obtained from crossing monosomic lines with interspecific monosomic F₁ hybrids: F₁: a – BC₁F₁(Mo11×F₁(766₃)) (494₂); b – BC₁F₁(Mo16×F₁(98₂)) (497₁); c – BC₁F₁(Mo19×F₁(769₄)) (8₈); d – BC₁F₁(Mo93×F₁(516₄)) (88₁) with chromosome 2 of the A_t subgenome of *G. barbadense*.

Univalents are indicated by arrows.

Pollen viability was assessed in BC₁F₁ monosomics via acetocarmine staining. Only two monosomic strains (922₈ and 923₇) presented high pollen viability (up to 94.21±1.06 %) (Fig. 4, Supplementary Material 12). Six monosomics methods resulted in a slight reduction in pollen viability (from 88.38±1.46 to 81.15±1.52 %), but five monosomics methods (7₂, 88₁, 88₄, 88₇, and 89₄) involving one variant of BC₁F₁(Mo93×F₁(516₄)) were characterized by a strong reduction in pollen viability (from 62.65±2.15 to 77.47±1.64 %).

Crossability, seed set, and germination characteristics of BC₂F₁ hybrid seeds with monosomic lines deficient in chromosome 2 of the A_t subgenome of *G. hirsutum*. Three monosomic lines deficient in chromosome 2 of the A_t subgenome of *G. hirsutum* from the cytogenetic collection were crossed with monosomic BC₁F₁(Mo×F₁(Mo×Pima 3-79)) hybrids that contained substitutions of chromosome 2 of the A_t subgenome of *G. barbadense*. Compared with those of the F₁ and BC₁F₁ hybrids, a strong decrease in crossability was observed (from 18.18 to 3.47 %) (Supplementary Material 13). A study of the seed set rate of hybrid BC₂F₁ plants revealed a significant decrease (from 47.47±5.02 to 27.69±5.55 %) compared with that of the F₁ and BC₁F₁ hybrids, while the germination rate of backcrossed BC₂F₁ seeds also decreased

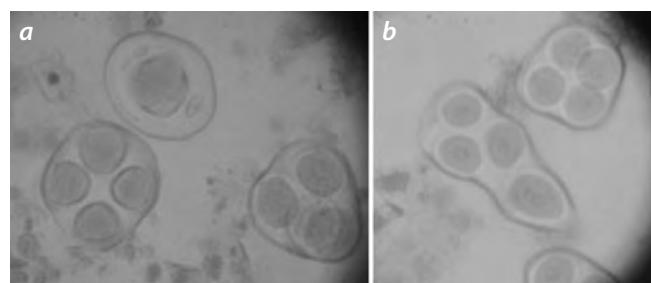


Fig. 3. Sporades in the hybrid plant BC₁F₁(Mo93×F₁(516₄)) (88₇): a – monad with three micronuclei and tetrads; b – abnormal tetrad and normal tetrad.

(from 69.23 to 44.44 %) compared with that of the same hybrids, with the monosomic line Mo93 exhibiting the strongest decrease in seed set and germination compared with the other two monosomic lines.

Identification of substitutions in the chromosome 2 of the A_t subgenome of *G. barbadense* in BC₂F₁ and BC₃F₁ hybrids via chromosome-specific molecular genetic markers. We previously demonstrated that five monosomic seedlings

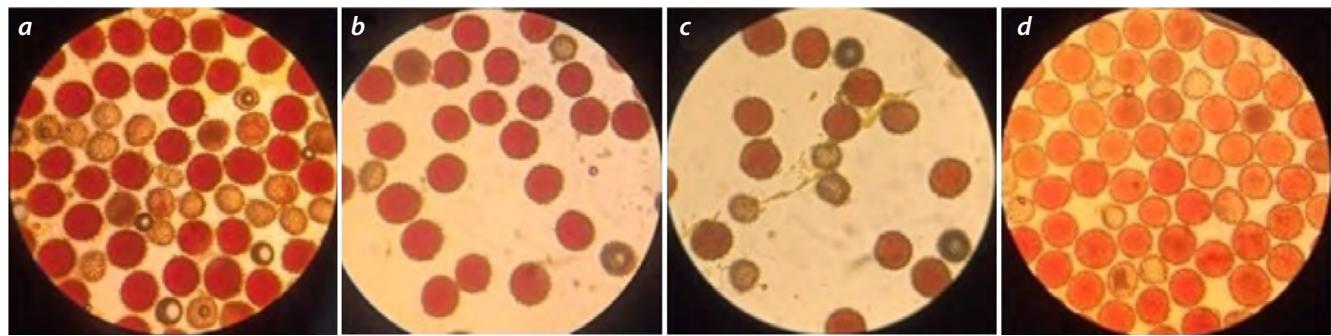


Fig. 4. Sterile (uncolored) and fertile (colored) pollen in monosomic BC₁F₁ hybrids obtained from crossing a monosomic line with a monosomic hybrid F₁(Mo₁₆×Pima 3-79): *a, b* (88₄) and *c, d* (89₄) in the BC₁F₁(Mo₉₃×F₁(516₄)) variant.

(21₁, 21₂, 21₄, 21₇ and 22₁) in the BC₂F₁(Mo₁₆×BC₁F₁(923₇)) variant with a putative substitution of chromosome 2 in the A_t subgenome of cotton were characterized by the presence of chromosome-specific alleles only from the L-458 line of *G. hirsutum*, whereas alleles from *G. barbadense* were absent, indicating the elimination of the alien chromosome (Sanamyan et al., 2023).

The results of a new study of two variants involving the Mo₁₆ line but two different BC₁F₁ hybrids (BC₂F₁(Mo₁₆×BC₁F₁(922₈))) and BC₂F₁(Mo₁₆×BC₁F₁(923₇))) revealed that in the first variant, three seedlings (817₁, 817₄ and 817₆) had chromosome-specific alleles only from the *G. barbadense* species, whereas alleles from the *G. hirsutum* species were absent; however, in the second variant, four seedlings (818₂, 818₃, 818₄ and 818₇) had alleles only from the *G. hirsutum* species, which indicated the elimination of the alien chromosome. Since previously reported chromosome-specific SSR markers, BNL3971, TMB1194, CIR376, and DPL0674 are located on chromosome 2 of the A_t subgenome of cotton (Liu et al., 2000; Gutiérrez et al., 2009; Saha et al., 2013, 2015; Wang et al., 2016), the obtained data indicate the presence of chromosome 2 substitutions in the first three seedlings (Fig. 5, Supplementary Materials 14, 19).

SSR-based deletion analysis in combinations BC₂F₁(Mo₁₉×BC₁F₁(88)) and BC₂F₁(Mo₉₃×BC₁F₁(88₄)) with putative substitution of chromosome 2 of the A_t subgenome of cotton allowed us to detect alleles of the *G. barbadense* species only in 10 backcross seedlings (820₂, 820₃, 820₇, 820₁₀, 820₁₄, 820₁₇, 821₂, 821₃, 821₆, and 821₁₀) of the first variant and in one (515₁) of the second, whereas alleles of the *G. hirsutum* species were absent. Chromosome-specific SSR markers, such as BNL834, BNL1434, BNL1897, BNL3292, BNL3413, BNL3424, BNL3547, BNL3971, BNL3972, JESPR101, JESPR179, JESPR304, TMB0471, TMB1194, CIR376, CIR381, CIR401, DPL0674, and NAU2277 are located on chromosome 2 of the A_t subgenome of cotton (Liu et al., 2000; Gutiérrez et al., 2009; Saha et al., 2013, 2015; Wang et al., 2016; https://www.CottonGen.org/data/download/marker_origin), and the obtained data indicated the presence of substitutions of this chromosome in the studied seedlings (Fig. 6, 7, Supplementary Materials 15–17, 19).

Similarly, in the BC₃F₁(Mo₉₃×BC₂F₁(515₁)) combination with a putative substitution of chromosome 2 of the A_t subgenome of cotton, alleles of *G. barbadense* were detected in

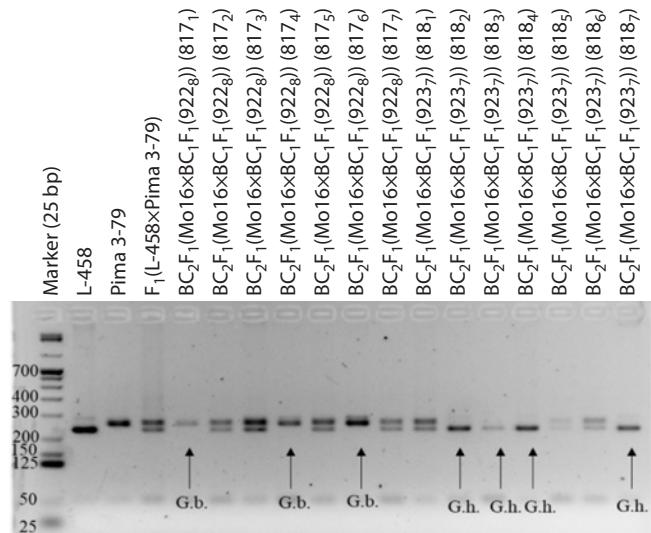


Fig. 5. Electropherogram of DNA amplicons of SSR markers in hybrid seedlings, BC₂F₁(Mo₁₆×BC₁F₁(922₈))) on chromosome 2 of the A_t subgenome of cotton.

Marker DPL0674.

seven backcrossed seedlings (839₁, 839₂, 839₄, 839₉, 839₁₀, 839₁₁, and 839₁₂), whereas alleles of the *G. hirsutum* species were absent. Because previously reported chromosome-specific SSR markers, DPL0674 and JESPR179, were located on chromosome 2 of the A_t subgenome of cotton (Liu et al., 2000; Gutiérrez et al., 2009; Yu et al., 2011; Saha et al., 2013, 2015; Wang et al., 2016), the obtained data indicated the presence of a substitution of this chromosome in the studied seedlings (Fig. 8, Supplementary Materials 18, 19).

Study of meiosis in BC₂F₁ hybrids with identified univalent. A study of chromosome pairing at the MI stage of meiosis revealed one backcrossing monosomic in each of the three backcrossing variants of BC₂F₁ (involving the Mo₁₆, Mo₁₉, and Mo₉₃ lines) at the time of writing.

Analysis of meiotic MI in three BC₂F₁ monosomic strains, including one monosomic strain, 21₁, from one cross with univalent chromosome 2 of the A_t subgenome of cotton *G. hirsutum* (Sanamyan et al., 2023) and two other monosomic strains (821₁₀ and 515₁) from two crosses with univalent

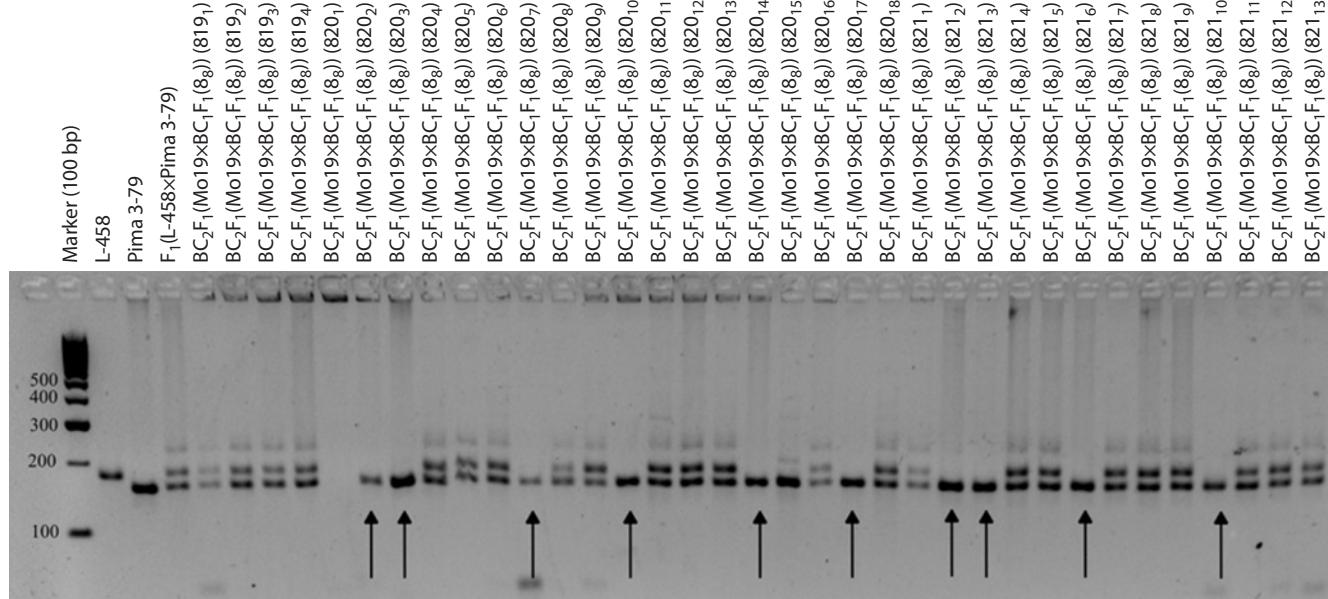


Fig. 6. Electropherogram of DNA amplicons of SSR markers in hybrid seedlings, BC₂F₁(Mo19×BC₁F₁(88₄)) on chromosome 2 of the A_t subgenome of cotton.

Marker JESPR179.

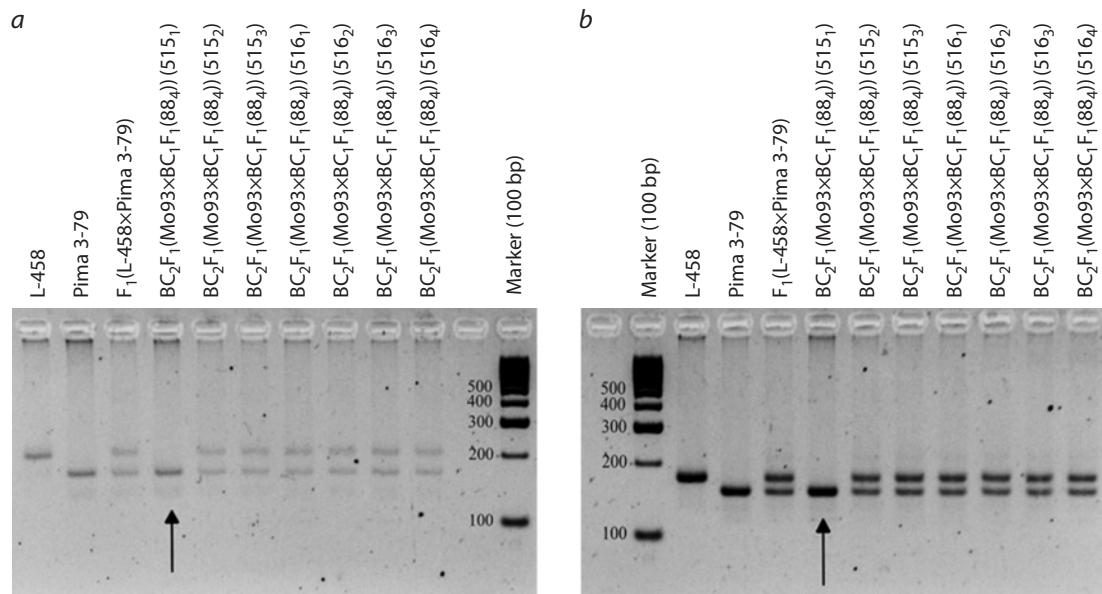


Fig. 7. Electropherogram of DNA amplicons of SSR markers in hybrid seedlings, BC₂F₁(Mo93×BC₁F₁(88₄)) on chromosome 2 of the A_t subgenome of cotton.

a – marker TMB0471; b – marker JESPR179.

chromosomes 2 of *G. barbadense*, revealed modal pairing of chromosomes with 25 bivalents and one univalent, confirming their monosomic status. Univalent size analysis in BC₂F₁ monosomics revealed a large chromosome 2 of *G. hirsutum* and a similar size in BC₂F₁ monosomic data with chromosome 2 of *G. barbadense*.

Characteristics of the transmission of chromosome 2 of the A_t subgenome of cotton during backcrossing of monosomic lines deficient in this chromosome by monosomic hy-

brids of different backcross generations. Molecular genetic analysis of backcrossed seedlings (at the stage of 3–5 true leaves), carried out via chromosome-specific molecular markers (SSR), allowed us to estimate the frequency and nature of the transmission of chromosome 2 from the A_t subgenome of cotton in BC₁F₁ hybrid plants. A study of five backcrossed BC₁F₁ families revealed that, in the BC₁F₁(Mo11×F₁(766₃)) variant, there was only one seedling (494₂) with the presence of the substituted chromosome, which indicated a very rare

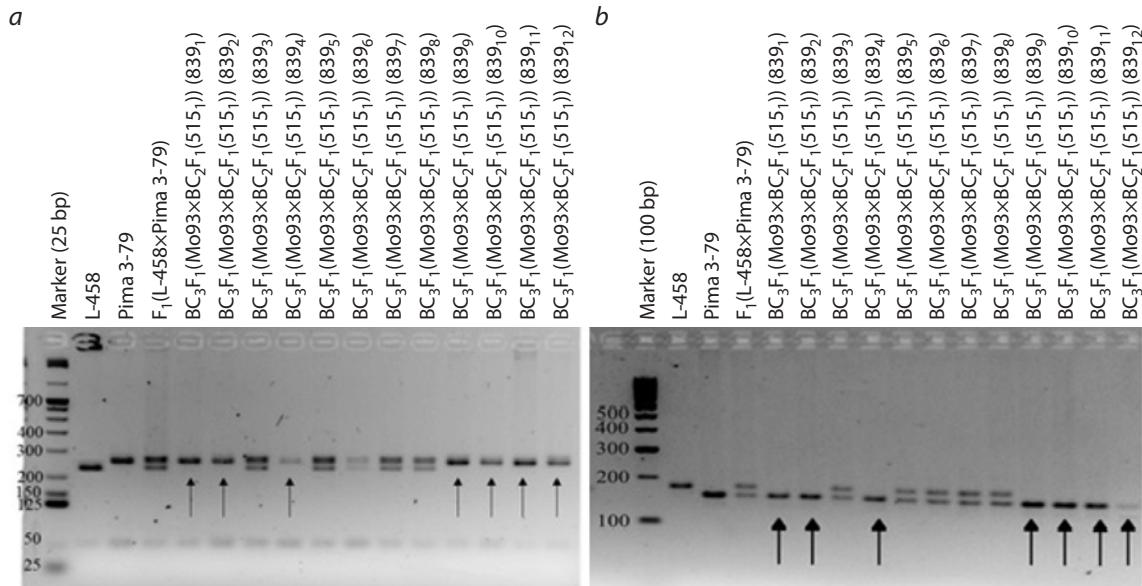


Fig. 8. Electropherogram of DNA amplicons of SSR markers in hybrid seedlings, BC₃F₁(Mo93×BC₂F₁(515₁)) on chromosome 2 of the A_t subgenome of cotton.

a – marker DPL0674; *b* – marker JESPR179.

transmission frequency of chromosome 2 of the A_t subgenome of the *G. barbadense* species (3.13 %) (Fig. 9, Supplementary Material 20).

Among the BC₁F₁ hybrids in the BC₁F₁(Mo16×F₁(98₆)) cross combination, seven seedlings with chromosome 2 substitutions were found, which were characterized by the presence of polymorphic alleles only from the *G. barbadense* species, indicating a relatively high frequency of transmission of chromosome 2 from the A_t subgenome of the *G. barbadense* species (21.21 %) in this variant. However, a study of the transmission of this chromosome in BC₂F₁ hybrids in the BC₂F₁(Mo16×BC₁F₁(923₇)) variant revealed the presence of chromosome-specific alleles only from the *G. hirsutum* species, whereas alleles from the *G. barbadense* species were absent, which revealed the absence of chromosome 2 substitution in five hybrids studied. Repeated analysis of BC₂F₁ hybrids in two cross variants allowed us to identify four hybrids in one BC₂F₁(Mo16×BC₁F₁(923₇)) variant with chromosome-specific alleles only from the *G. hirsutum* species, which drew attention to the repeated elimination of this chromosome, whereas in the other variant (922₈), the presence of chromosome 2 substitution from the *G. barbadense* species was observed. In general, in BC₂F₁ with the participation of the monosomic line Mo16, the frequency of transmission of the substituted chromosome was characterized by a decrease (up to 12.00 %) compared with that in BC₁F₁ hybrids, as well as elimination in the majority of hybrids (36.00 %). Moreover, elimination of the substituted chromosome occurred in one variant of crosses involving the same BC₁F₁ hybrid plant, which suggested the influence of genotype on the nature of chromosome transmission (Fig. 9, Supplementary Material 20).

Among the hybrids of one family in the BC₁F₁(Mo19×F₁(769₄)) variant with a putative chromosome 2 substitution, a single hybrid plant (8₈) was identified that had an allele from *G. bar-*

badense, which indicated the transmission of the substituted chromosome 2 with a low frequency (12.50 %). However, when the transmission of this chromosome in BC₂F₁ hybrids in the BC₂F₁(Mo19×BC₁F₁(8₈)) variant was studied, chromosome-specific alleles from the *G. barbadense* species were detected only in ten hybrids in three analysed families, which indicates greater transmission of the substituted chromosome 2 (28.57 %) than in BC₁F₁ hybrids (Fig. 9, Supplementary Material 20).

Nine hybrids with chromosome 2 substitutions were detected in the BC₁F₁(Mo93×F₁(516₄)) variant, which was characterized by the presence of alleles from *G. barbadense* only, indicating a high frequency of transmission of this chromosome (42.86 %). Among the BC₂F₁ hybrids in the BC₂F₁(Mo93×BC₁F₁(88₄)) variant, only one hybrid from two families had a substitution from *G. barbadense*, confirming the low frequency of chromosome 2 substitution (14.29 %). However, the study of the transmission of this chromosome in BC₃F₁ hybrids in the BC₃F₁(Mo93×BC₂F₁(515₁)) variant made it possible to characterize the presence of chromosome-specific alleles from the *G. barbadense* species in seven hybrids out of 12 studied, i. e., the transmission of chromosome 2 was observed in more than half of the hybrids (58.33 %) (Fig. 9, Supplementary Material 20).

Morphobiological analysis of monosomic hybrids BC₁F₁, BC₂F₁, and BC₃F₁ obtained from crosses of the monosomic cotton lines *G. hirsutum* with chromosome 2 deficiency of the A_t subgenome with monosomic hybrids of different generations of backcrosses with chromosome substitution. Morphobiological analysis of backcross monosomic hybrids BC₁F₁ with chromosome 2 substitution of the A_t subgenome of cotton revealed short stature (up to 50 cm on average); reduced size of three-lobed leaves, buds, and flowers; low foliage; shortened internodes; and weak budding and flowering during the first year of vegetation (only 2–3 buds

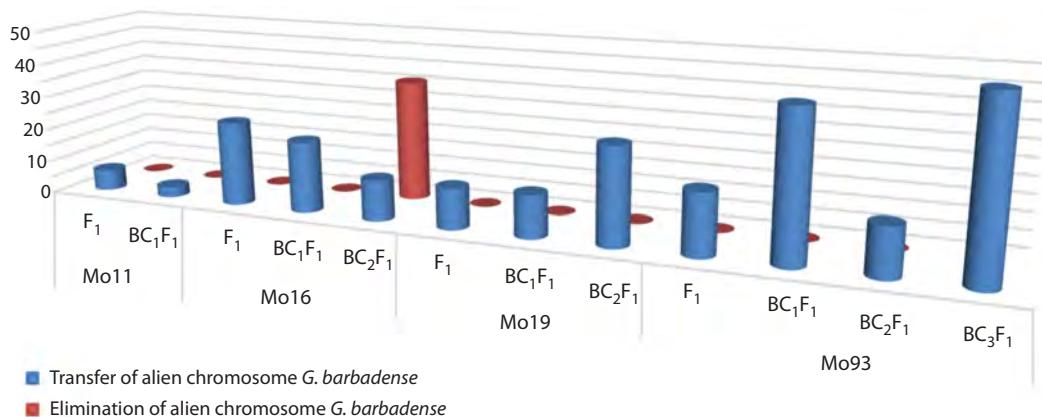


Fig. 9. Frequency of occurrence of monosomic hybrids with alien chromosome 2 of the A_t subgenome of cotton *G. barbadense*.



Fig. 10. Plants of the original lines and monosomic hybrids of cotton F₁ and BC₂F₁ obtained from crossings of the recurrent parent with monosomic hybrids.

a – monosomic line Mo16 for chromosome 2 of the A_t subgenome; b – F₁(Mo16×Pima 3-79) (98₂); c–e – BC₁F₁(Mo16×F₁ 98₆) with substitution of chromosome 2 of the A_t subgenome of cotton *G. barbadense*; c – 923₇, d – 495₅, e – 497₄; f – BC₂F₁(Mo16×BC₁F₁ 922_g) (817₄) with substitution of chromosome 2; g – BC₂F₁(Mo16×BC₁F₁ 923₇) (21₁) without substitution of chromosome 2 of the A_t subgenome of cotton *G. barbadense*.

and flowers per plant) compared with the recurrent parent. The backcross hybrids with the participation of the monosomic line Mo16 (Fig. 10) were characterized by particularly short stature and a decrease in growth rates, whereas in the second and third years of vegetation, there was an increase in the size of the leaves, the number of buds and the number of opened bolls.

Compared with the monosomic hybrid BC₁F₁, with the substitution of chromosome 2, the complemented hybrids BC₂F₁ and BC₃F₁, with the substitution of chromosome 2 of the A_t subgenome of cotton, were distinguished by taller growth (up to 80 cm), medium foliage, larger three-lobed leaves, an increased number of flowers and medium-sized spherical bolls.

Discussion

Interspecific crosses of various tetraploid cotton species are characterized by high crossability and fertility of first-generation hybrids. However, in later generations, decreases in all these parameters and mass sterility are observed (Mauer, 1954; Abdullaev, 1974).

A study of the crossability and seed set of backcrossed seeds obtained from crosses of monosomic lines deficient in chromosome 2 of the A_t subgenome of cotton *G. hirsutum* with interspecific monosomic F₁ hybrids and backcrossed monosomic BC₁F₁ hybrids revealed a significant decrease in crossability in all the studied backcrossed hybrids, whereas hybrid seed set and germination decreased in most hybrids. This decrease was explained by the decrease in these para-

meters in the original monosomic lines due to the presence of a significant number of nullisomic gametes, as well as the hemizygosity of the maternal and paternal plants, which resulted in the presence of numerous unfertilized eggs in the form of motes and low germination of nullisomic seeds.

Molecular genetic methods using microsatellite sequence markers (SSRs), which are widespread in eukaryotic genomes and exhibit a high level of polymorphism, are widely used to identify alien introgression. To date, several collections of such markers have been created for cotton (BNL, JESPR, CIR, DPL, and NAU), and molecular genetic linkage maps have been constructed due to chromosomal specificity of many SSR markers (Liu et al., 2000; Gutiérrez et al., 2009; Yu et al., 2011; Saha et al., 2013, 2015; Wang et al., 2016; https://www.cottongen.org/data/download/ marker_origin). The presence or absence of microsatellite marker amplification products after polymerase chain reaction allows us to judge the presence or absence of an alien chromosome in the studied genotype. Therefore, our molecular genetic screening of backcross hybrids at the seedling stage allowed us to quickly identify the substitution of chromosome 2 of the A_t subgenome of cotton of the *G. barbadense* species.

Compared with direct cytological observation, PCR-based screening for chromosome-specific markers has been shown to be more productive and significantly more effective (Polgári et al., 2019). This is especially true when it is impossible to detect and monitor alien genetic material directly on cytological preparations via FISH and GISH hybridization.

The originality of our approach lies in the development of a new scheme for the targeted, planned substitution of specific chromosomes via monosomic lines with previously established univalent chromosome identities and chromosome-specific microsatellite markers (SSRs) at the seedling stage. During the creation of new chromosome substitution lines via this scheme, significant differences were identified between specific chromosomes in the cotton genome in terms of the frequency of transmission of alien chromosomes, their elimination at different stages of backcrossing, and the morphobiological characteristics of the backcrossed hybrids. This study presents data on the specific consequences of the transfer of genetic material from chromosome 2 of the A_t subgenome of cotton *G. barbadense* into the genome of cotton *G. hirsutum* via four monosomic lines.

The results of the study of backcross progenies BC₁F₁, BC₂F₁ and BC₃F₁ (in one variant of crossing) revealed that the frequency of transmission of chromosome 2 of the A_t subgenome of cotton, *G. barbadense* depended on both the genotype of the monosomic line and the genotype of the backcross hybrid used in the crossing. Thus, the level of transmission of chromosome 2 of the A_t subgenome of *G. barbadense* in the first backcross generation, with the participation of four monosomic lines, varied within a wide range (from 3.13 to 42.86 %), whereas in the BC₂F₁ generation, where only three monosomic lines were involved, the transmission frequency was noticeably reduced and varied within a narrower range (from 12.00 to 28.57 %). However, in the BC₃F₁ generation, which included only one monosomic line (Mo93), the highest frequency of chromosome 2 transmission (58.33 %) was observed in more than half of the studied hybrids. Moreover,

this line was characterized by the highest average number of substitutions per hybrid genome among all four studied lines. In addition, the elimination of chromosome 2 of the A_t subgenome was detected in the BC₂F₁ variant (Mo16×BC₁F₁ (923₇)), which involved the same paternal backcross monosomic hybrid plant (923₇) during two experiments, indicating the preferential elimination of chromosome 2 in the studied backcross progenies and the influence of a specific paternal genotype on this process. Therefore, the analysis of the transmission characteristics of alien chromosome 2 revealed differences in the competitiveness of this chromosome when it is transmitted to offspring in the genotypic environment of four monosomic lines.

The success of substitution depends on how well the alien chromosome compensates for the missing chromosome, since it is difficult to assume that each alien chromosome could compensate equally for the absence of homeologous chromosomes (Morris, Sears, 1970).

Previously, a dependence of rye chromosome introgression on the genotypic environment of the recipient was discovered, since differences in the frequency of rye chromosomes were observed in the F₂ hybrid population between a wheat-rye substitution line for chromosomes 1R+2R and the winter wheat varieties Holme and Kraka (Merker, Forsstrom, 2000). Notably, the genotype of the variety also influenced the frequency of telocentric formation on the rye chromosome and T2R.2DL translocation (Krasilova et al., 2011).

Cases of complete uniparental elimination of chromosomes from the entire genome are widely known (Ishii et al., 2016; Dedukh, Krasikova, 2021). They result from hybridization between distantly related species as an element of protecting the integrity of the genome from “genomic shock” (McClintock, 1984). An example is the preferential elimination of chromosomes from the entire D genome in first-generation hybrids during wheat-rye hybridization (Li et al., 2015).

Examples of partial elimination of individual chromosomes are less numerous, but it is known that in each specific case, different mechanisms of chromosome elimination operate in interspecific hybrids, depending on the specific species involved. Thus, an assessment of a large population of wheat-barley hybrids via genomic *in situ* hybridization (GISH) and simple sequence repeat (SSR) markers revealed the absence of preference for the elimination of individual barley chromosomes compared with wheat chromosomes (Polgári et al., 2019). A study of the transmission of chromosome 5R through the gametes of wheat-rye dimonosomics 5R5D revealed a significantly lower competitive ability of this chromosome in transmission to offspring and its preferential elimination (Silkova et al., 2011). Selective elimination of chromosomes in *Hordeum bulbosum* L. is associated with the loss of one of the types of histone protein H3 (CENH3) in the centromere, leading to its inactivity and absence of chromosome attachment to the mitotic spindle, as well as the formation of micronuclei and their degeneration (Sanei et al., 2011). A comparative analysis of the nucleotide sequences of the centromeric histone CENH3 genes in wheat-rye allopolyploids of different ploidy levels revealed increased expression of rye CENH3 variants, which is associated with the maintenance of a viable hybrid genome (Evtushenko et al., 2019). Using bread wheat as an example,

a wide range of features of alien chromatin introgression was demonstrated, which represents significant potential for gene pool enrichment (Adonina et al., 2021).

Although screening for the presence of alien chromosomes in backcrossed cotton progenies via molecular genetic markers made it possible to detect specific consequences of the introgression of chromosome 2 of the A_t subgenome of the *G. barbadense* species at different stages of backcrossing, the study of the behavior of univalent chromosomes at the MI stage of meiosis revealed the similarity of pairing in backcrossed monosomes with the univalent chromosome 2 of the A_t subgenome of the cotton species *G. hirsutum* and the univalent chromosome 2 of the A_t subgenome of the *G. barbadense* species.

The backcrossed BC₁F₁ monosomic strains we examined were characterized by a general delay in the development of vegetative and generative organs, manifested by stunted growth, reduced foliage, and poor budding and flowering during the first year of vegetative development. This hampered their study and backcrossing and significantly complicated and delayed the creation of chromosome substitution forms. However, during the second year of vegetative development, the backcrossed plants showed normalization of vegetative and generative organ development.

Similar dwarfism was observed in the monosomic alien complemented cotton line *G. hirsutum*/*G. bickii* Proch. with a 10 Gb chromosome substitution (MAAL), created through crosses with an amphidiploid (2n = 78, AADDG₁G₁) and chromosome-specific SSR markers (Tang et al., 2018). It can be assumed that such changes in morphobiology occurred due to hybrid genome reorganization and introgression of alien chromatin.

Conclusion

This study demonstrated the negative consequences of the introgression of chromosome 2 of the A_t subgenome of cotton, *G. barbadense*, into the genome of cotton, *G. hirsutum*, involving four monosomic lines. These negative consequences include decreased crossability, seed set, and germination of hybrid seeds; a wide variation in the transmission level of alien chromosome 2 (from 3.13 to 42.86 %) in BC₁F₁, but a less narrow one (from 12.00 to 28.57 %) in BC₂F₁; elimination of chromosome 2 of the A_t subgenome of the *G. barbadense* species in the BC₂F₁(Mo16×BC₁F₁(923₇)) variant with the same paternal genotype, which indicated the influence of a specific paternal genotype on this process; dwarfism; and reduced foliage, weak budding and flowering in the first year of vegetative development but an increase in the number of buds, flowers and bolls in monosomic hybrids of the following backcross generations.

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